

# *Molar Pregnancy and Gestational Trophoblastic Disease*

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# Conflict of Interest Disclosure

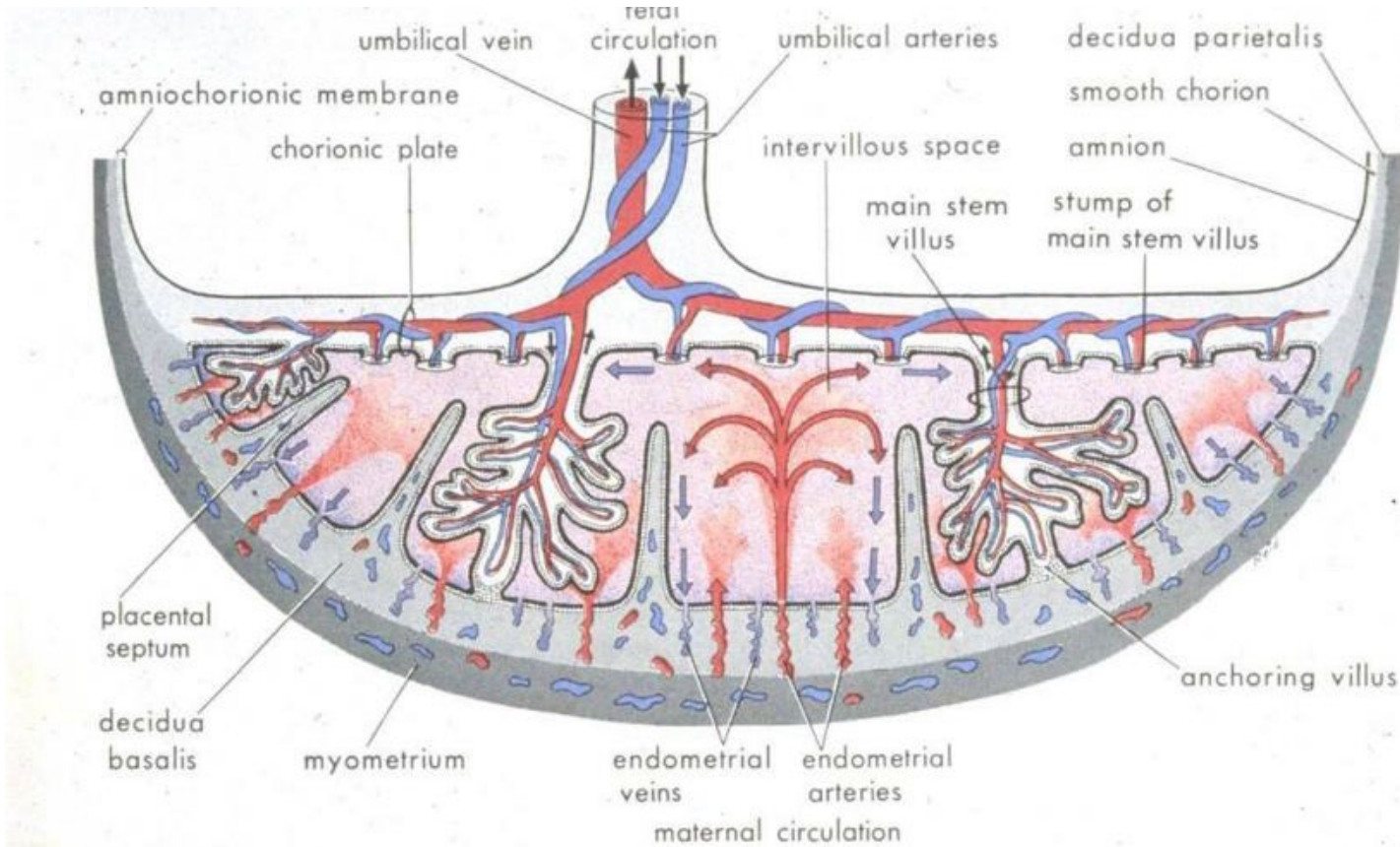
*None*

# Goals

- **Molar Pregnancy and Gestational Trophoblastic Disease**
  - Highlight molar gestations in their pathobiological context
  - Define the histologic and genetic features that distinguish molar gestation from each other and their mimics
  - Provide a practical strategy for pathologic diagnosis that leverages your available ancillary tools
  
- ***Breaking News: COVID-19 in Pregnancy***
  - Review the pathobiology of SARS-CoV-2 infection in the placenta and its clinical implications
  - Define the features of COVID-2 placentitis



# Molar Pregnancy – Key Concepts



- Products of conception
- Genetically distinct from the host in which they reside
- Arise most frequently from genetic errors occurring in fertilization
- Like normal trophoblasts, invade tissue and blood vessels
- Nosologic organization begins with presence or absence of villi
- Phenotypes resemble cells constituting the normal placenta
- Must be considered in the spectrum of trophoblastic neoplasia

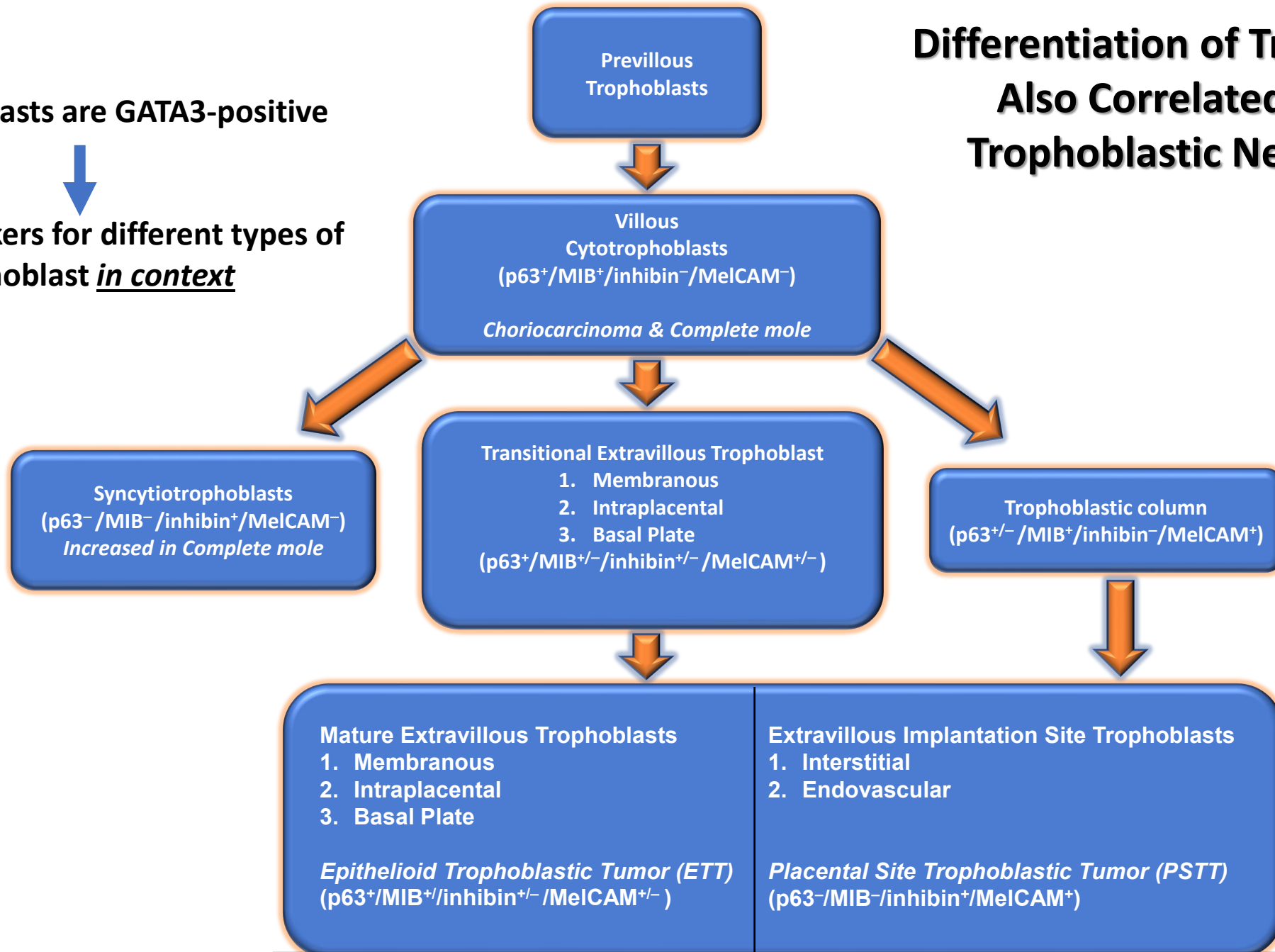


All trophoblasts are GATA3-positive



Specific markers for different types of trophoblast *in context*

## Differentiation of Trophoblast Also Correlated With Trophoblastic Neoplasia



# Nosology Of Trophoblast Neoplasia

Tumor	Fetus	Cyto-trophoblast	Syncytio-trophoblast	Intermediate (Implantation Site) Trophoblast	Chorionic Membrane Trophoblast	Villi	Biological Potential	Primary Differential Diagnosis	Typical Genetics
Partial Hydatidiform Mole	✓	✓	✓	✓	✓	✓	?	Aneuploidy (+18), Complete Mole +/- co-twin, Mesenchymal Dysplasia, Beckwith-Wiedemann Syndrome	3n with diandry (2p1m)
Complete Hydatidiform Mole		✓	✓	✓		✓	Invasive & Metastatic	Hydropic Abortus, Partial Mole	2n with diandry (2p)
Chorio-carcinoma	✓	✓	✓				Invasive & Metastatic	Other High Grade Neoplasm	Diploid, possibly diandric
Placental Site Trophoblast Tumor (PSTT)	✓			✓			Invasive & Metastatic to Lungs, pelvis, LNs (30-50%)	"Exaggerated" & Molar Implantation	46, <u>XX</u>
Epithelioid Trophoblast Tumor	✓				✓		Invasive	SCC of cervix ?De-diff. C	46, <u>XX</u>

# Risk Factors For Trophoblastic Neoplasia

- Maternal Age
  - < 20 y/o RR = 1.5
  - >40 y/o RR = 5.2
- Race/Ethnicity
  - Indonesia Incidence = 1:85
  - Japan Incidence = 1:522
  - Sweden Incidence = 1:1560
  - USA Incidence = 1:1724
  - Asian women in the USA have a higher risk than other ethnic groups
- Prior Molar Pregnancy, especially >1
- Nulliparity
- Low socioeconomic status

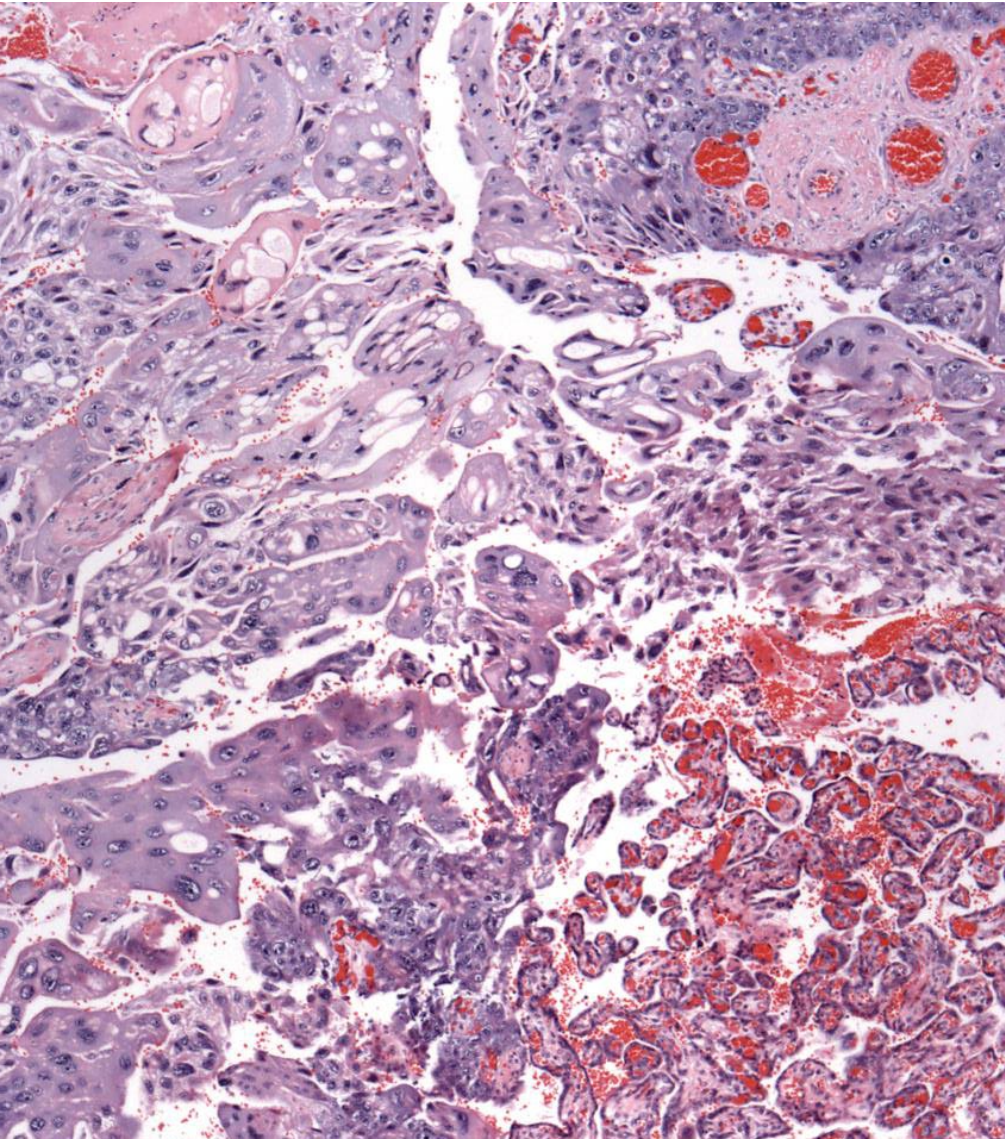
(best studied in complete hydatidiform mole)



# Choriocarcinoma (ChorioCA)

## The Foundation Of Trophoblastic Neoplasia



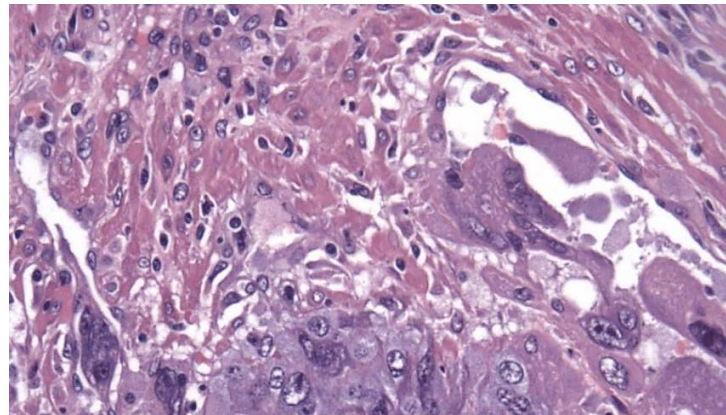
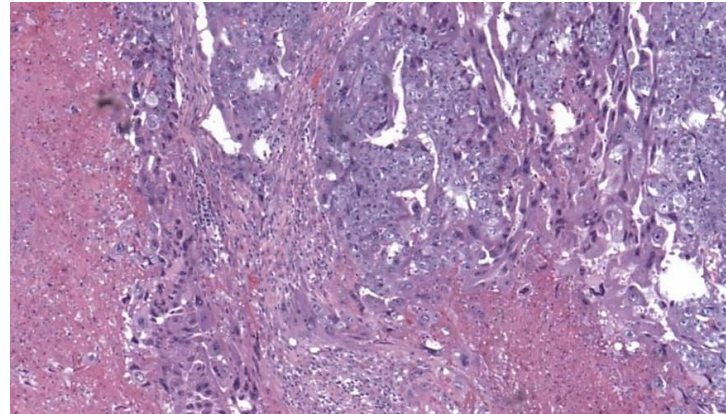
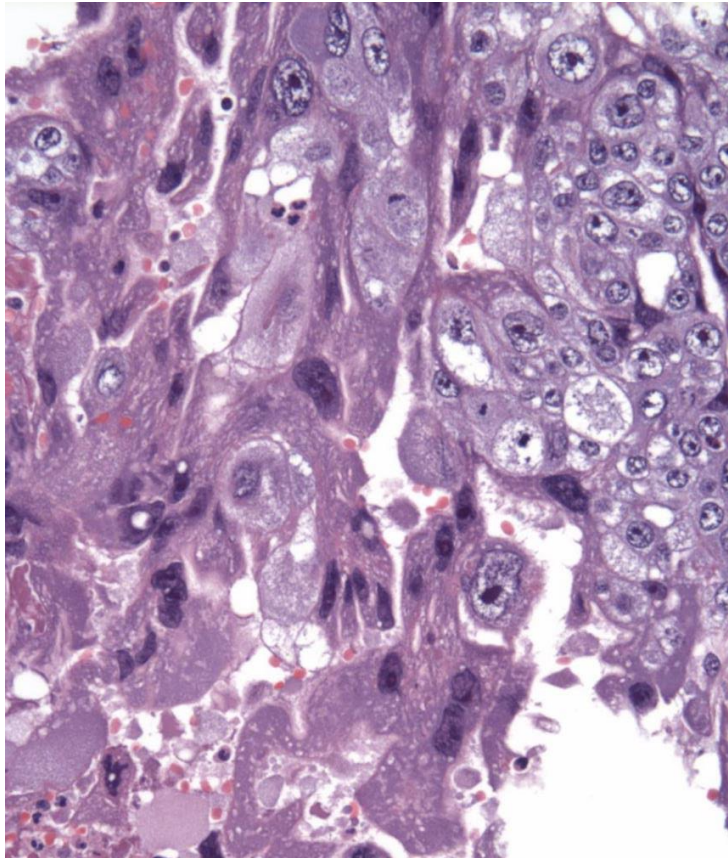


# Choriocarcinoma – Clinical Context

- Gestational association:
  - 50% molar (complete >> partial)...but only 1 in 40 molar pregnancies
  - 25% spontaneous abortion
  - 2.5% ectopic gestation
  - 22.5% normal pregnancies (most often in 3<sup>rd</sup> trimester)
- Time Interval
  - Late -- many years (report of 14 yr)
  - Early -- concurrent (intraplacental choriocarcinoma)

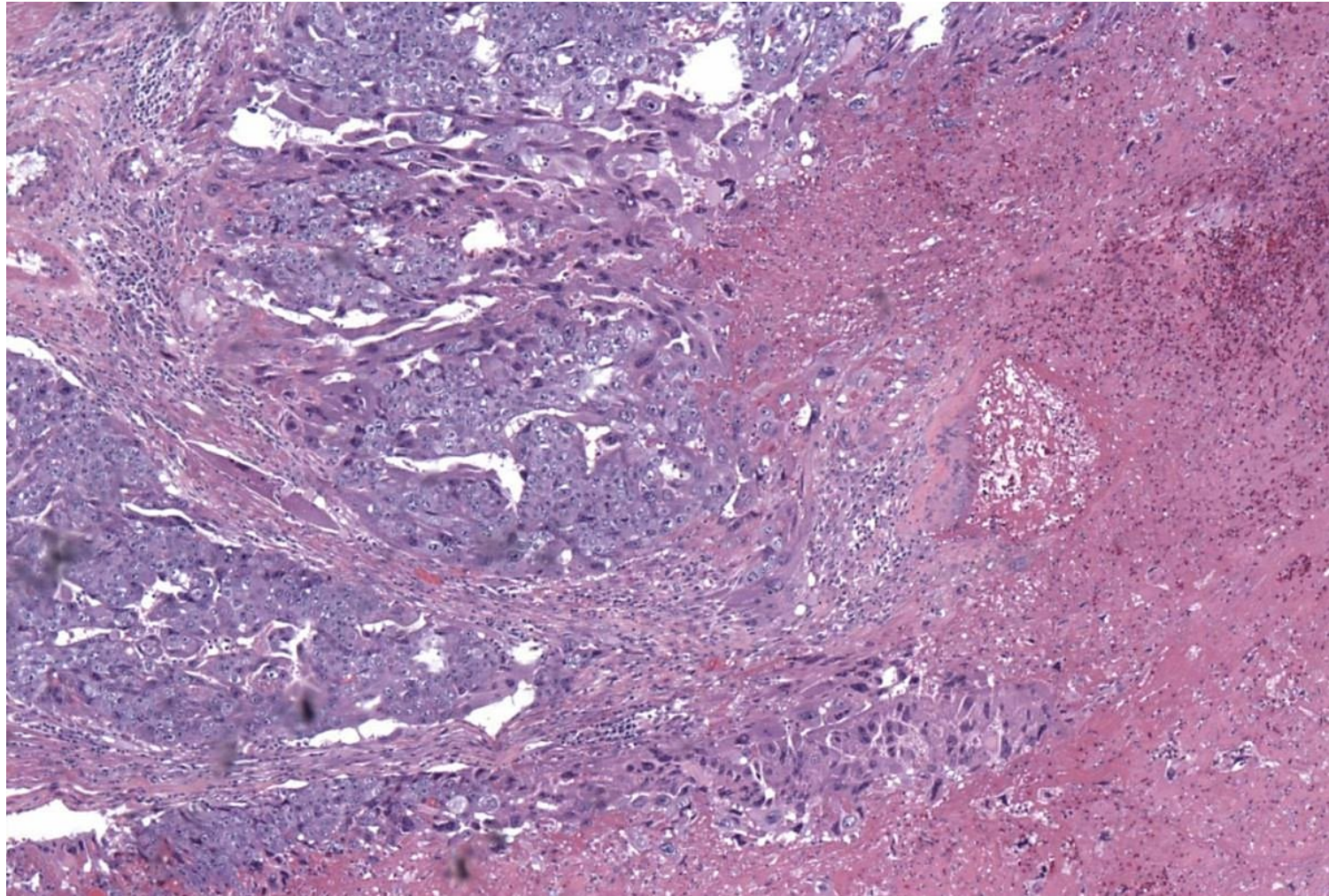


# Choriocarcinoma- Diagnosis



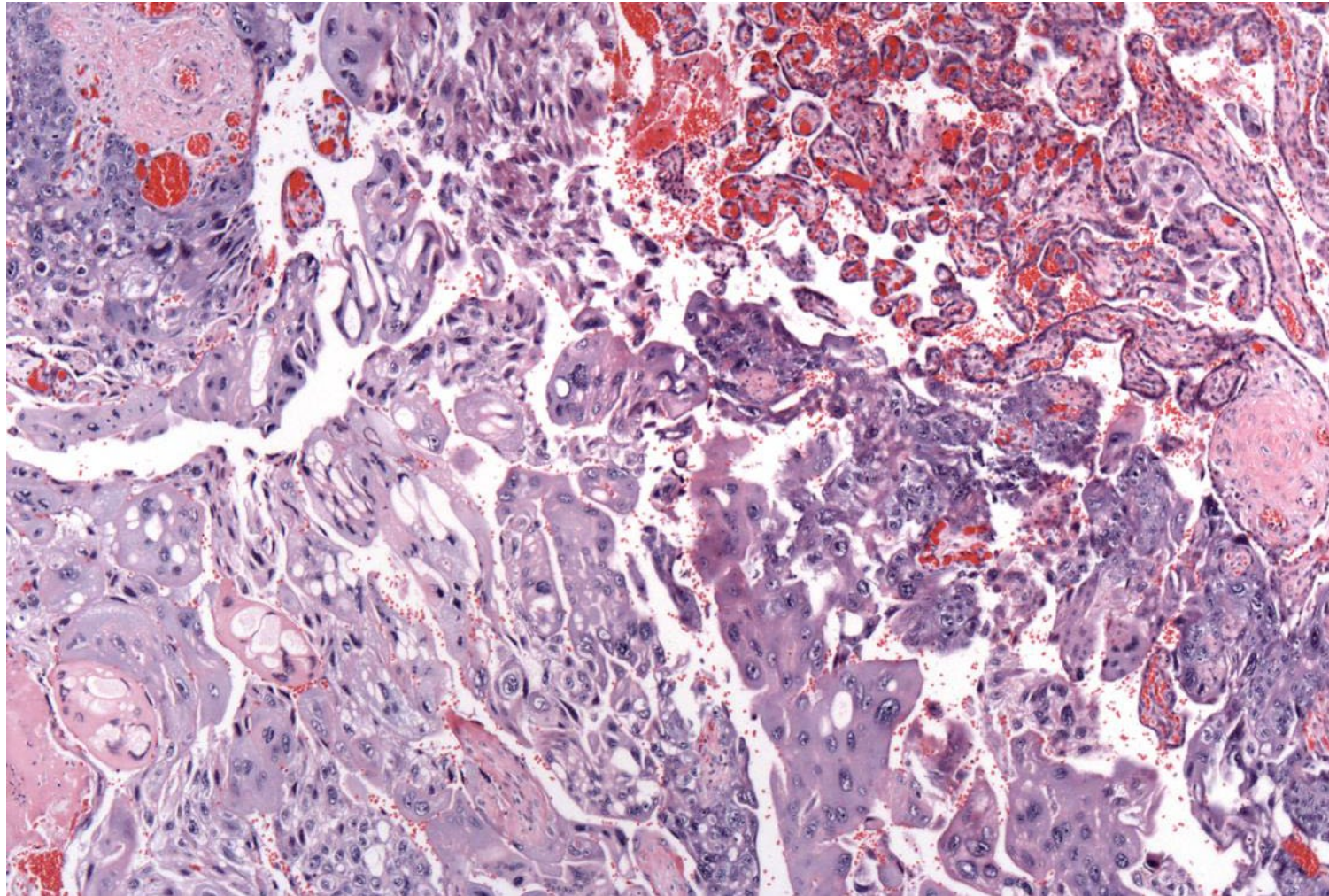
- **Gross features**
  - Hemorrhagic nodule(s)
  - Sampling of blood clot...particularly at the periphery
- **Histological features**
  - Biphasic (all cytokeratin +, GATA3)
    - Cytotrophoblasts and intermediate trophoblasts
    - Syncytiotrophoblasts (strong + for  $\beta$ -HCG, weak for HPL)
      - **Caveat: may be infrequent in some tumors, possibly reduced following chemoTx**
  - Hemorrhage and necrosis may predominate





Choriocarcinoma  
with  
hemorrhagic  
necrosis

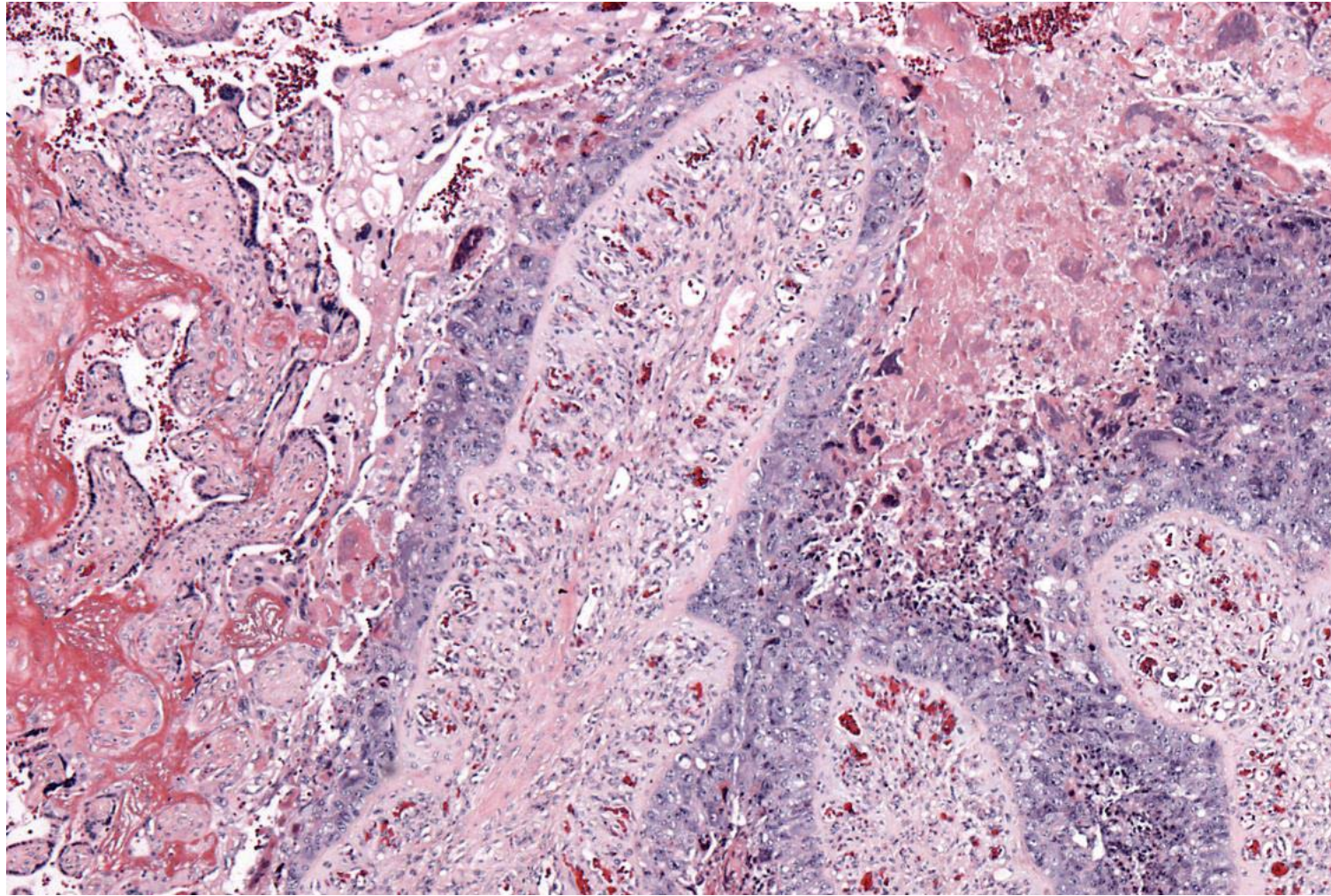




Choriocarcinoma  
During  
Pregnancy:

Intraplacental  
choriocarcinoma  
in term non-  
molar placenta





Intraplacental  
choriocarcinoma  
in term non-  
molar placenta

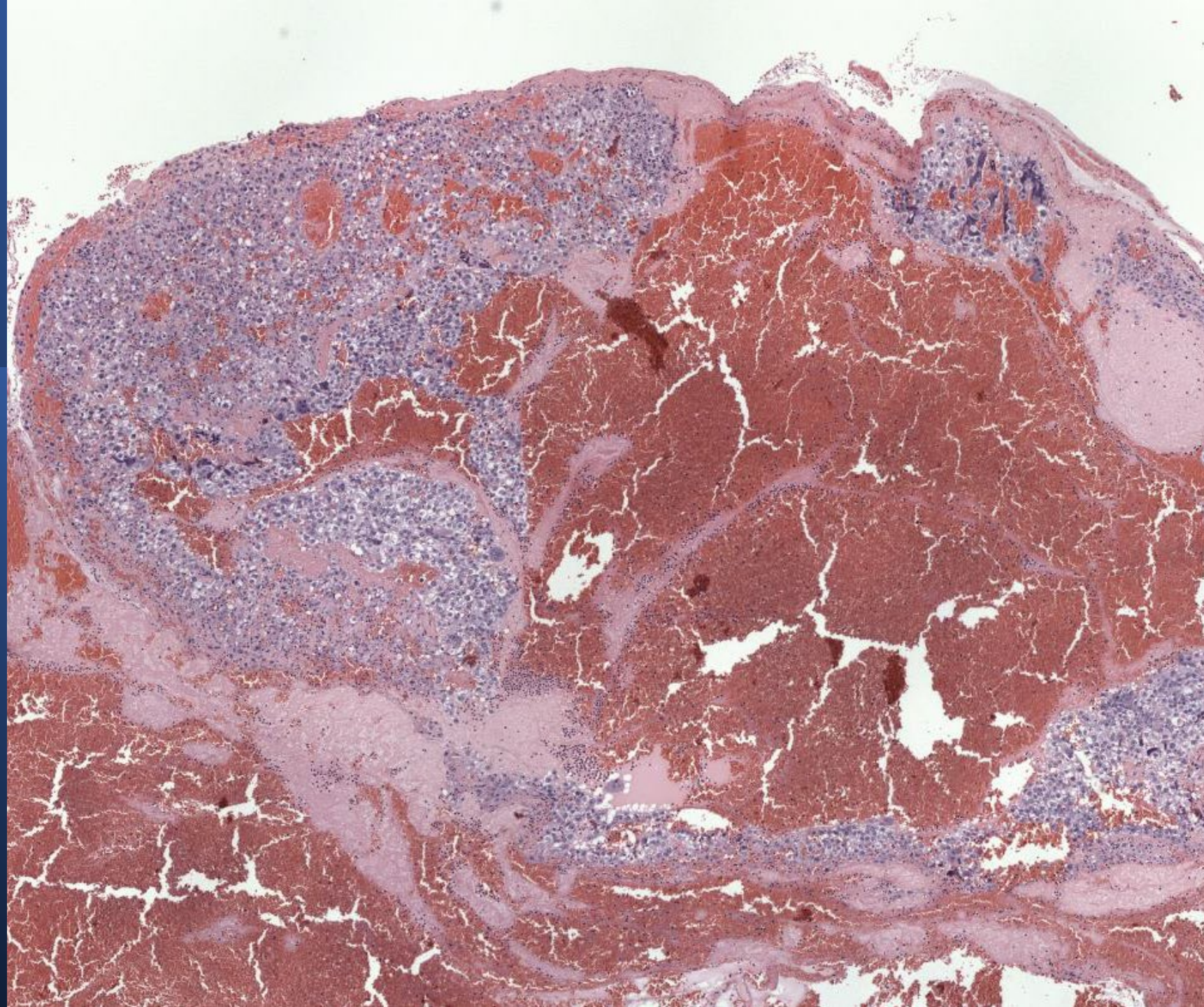


# Choriocarcinoma

- **Differential Diagnosis:**
  - Pre-villous trophoblasts in early gestations
  - Persistent Trophoblastic Disease following Complete Mole
    - No villi due to sampling
    - Residual molar implantation site
    - Choice of terminology does not have clinical impact
  - Placental Site Trophoblast Tumor (PSTT) or Epithelioid Trophoblast Tumor (ETT)
    - “Syncytiotrophoblast-poor” choriocarcinoma



Mimic of  
Choriocarcinoma:  
  
Early gestation with  
trophoblast shell in  
hematosalpinx/ectopic  
pregnancy





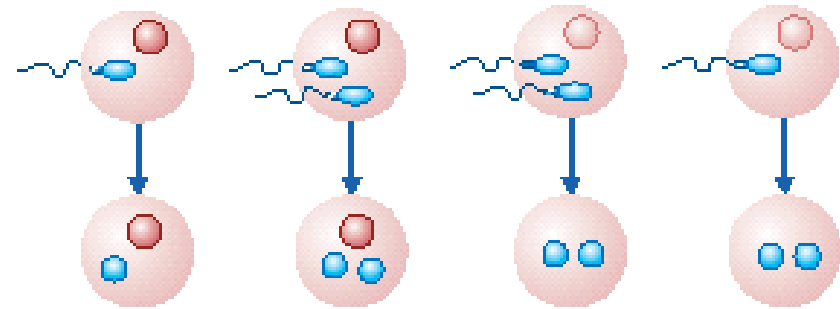
# Complete Hydatidiform Moles (CHM)





# Classic Complete Hydatidiform Mole – Genetics

- Diploid with diandry (2p)
  - p57 is absent because it is expressed only from the maternal genome...in cytotrophoblasts and stromal cells



	Normal	Partial mole	Complete hydatidiform mole	
Ploidy	2n	3n	2n	1n>2n (duplication)
Parents	Biparental	Biparental	Paternal	Paternal
Chorio Ca risk	Low	Low	Elevated	Elevated

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20%	80%
46,XX or XY	46,XX
Genome-wide heterozygosity	Genome-wide LOH

# Biparental Complete Hydatidiform Mole

- CHM variant that are ***diploid but biparental (1M:1P)***
- Recurrent
- Autosomal Recessive
- Maternal effect -- failure to establish maternal imprint
- Specific trans-acting genes:
  - *NALP7/NLRP7* (MIM 609661)
    - 19q13.42
    - Negative regulator of interleukin-1- $\beta$ , which may in turn regulate inflammatory proteases needed for blastocyst implantation
  - *C6orf221* (MIM 611687)
    - 6q13
    - aka Embryonic Stem Cell-associated Transcript 1; ECAT1
    - Member of eutherian oocyte- and embryo-expressed KHDC1/DPPA5/ECAT1/OOEP gene family at 6q13
    - Oocyte specific expression

# Complete Hydatidiform Mole

- As the gestational age increases, the molar villi uniformly grow to macroscopic size, forming grape-like cysts. This entity is so named because of the “water drop” cysts (Greek - “hydatisia”) cluster together forming a false conception (Latin “mole”).
- Embryonic development does not occur.

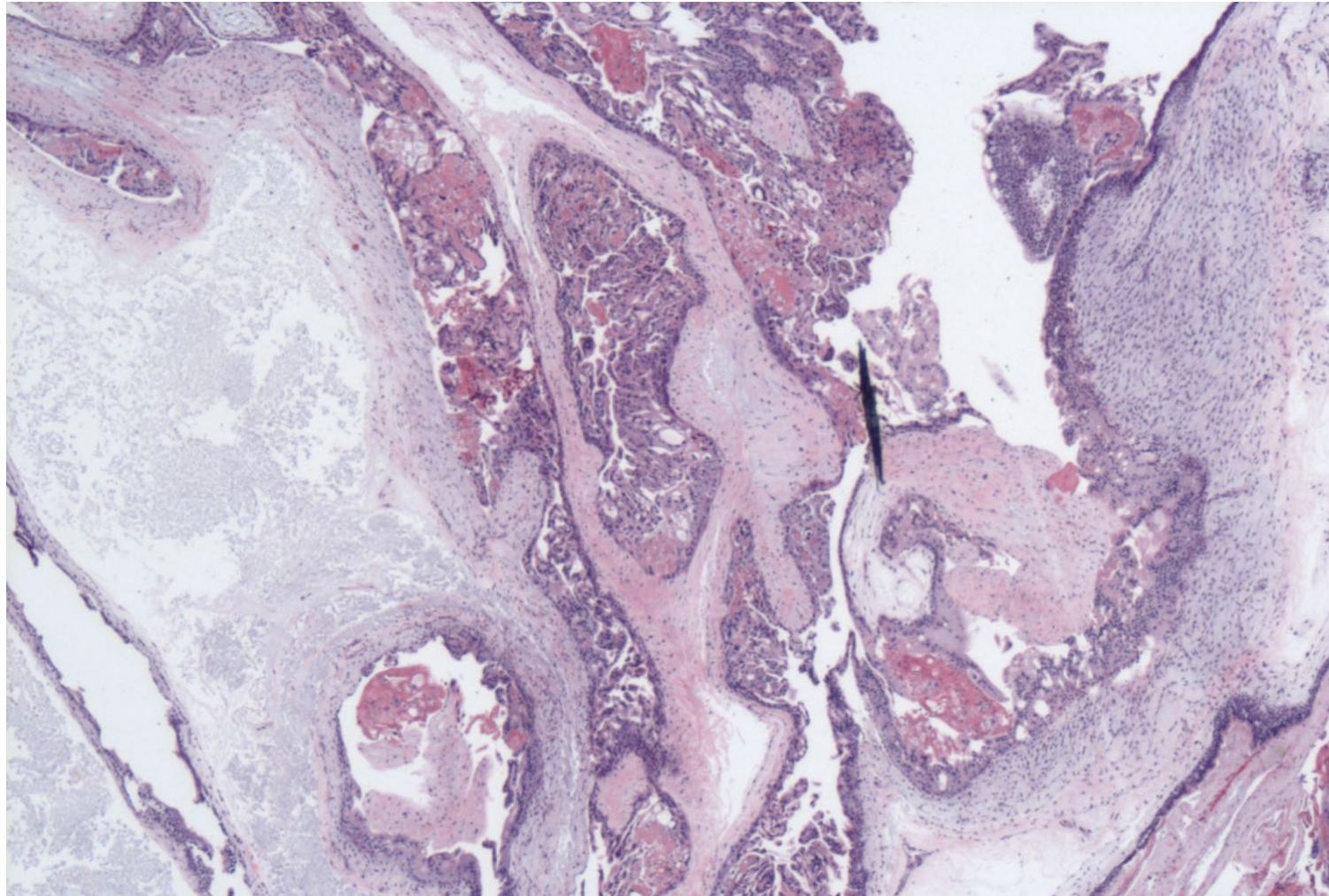




# Complete Hydatidiform Mole

## Histological Features

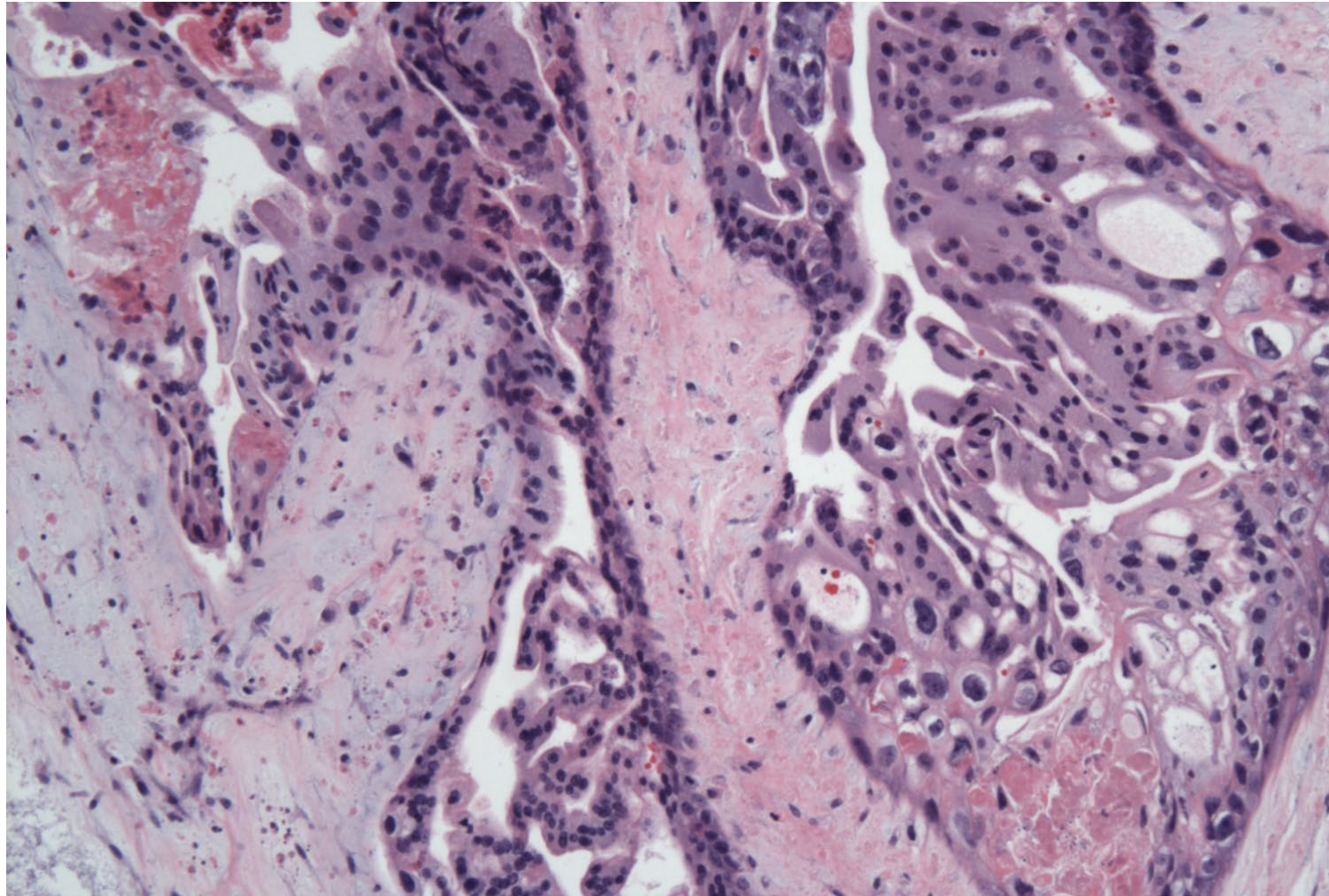
- Complete mole  $\approx$  Choriocarcinoma plus villi
- Trophoblast atypia (usually)...yes, even more than usual
- Circumferential trophoblast hyperplasia
- Molar (atypical) implantation site
- Histological features evolve with time
  - Blue myxoid stroma, similar to early normal gestations
  - Villi have narrow cleft-like invaginations to the stroma
  - Progressive stromal cell death degeneration
  - Vessels without blood may be present very early
  - Then cavitation of villous stroma



## Complete Hydatidiform Mole

- **Cavitation**
- **Absence of fetal vessels**

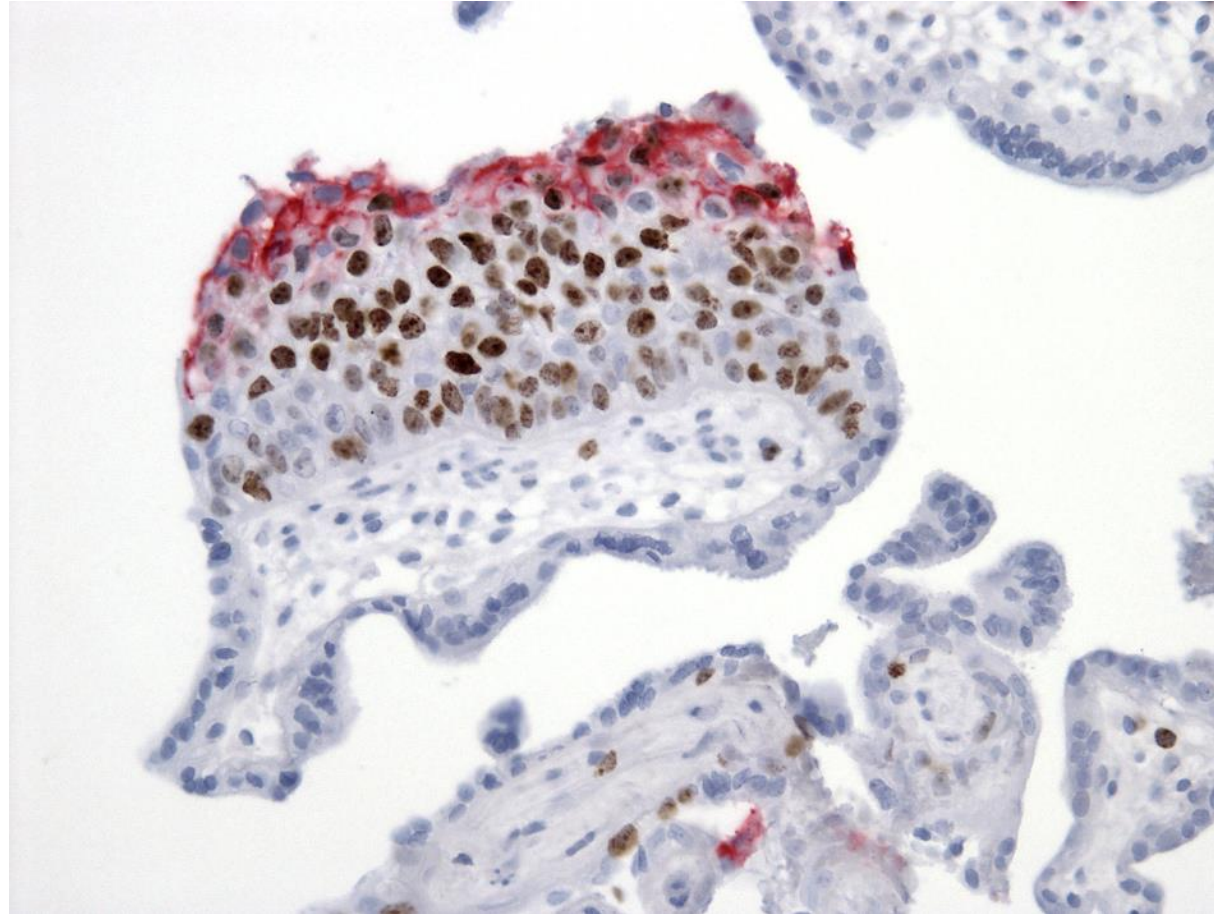




## Complete Hydatidiform Mole

- **Trophoblast hyperplasia**
  - **Circumferential**
  - **Exuberant**

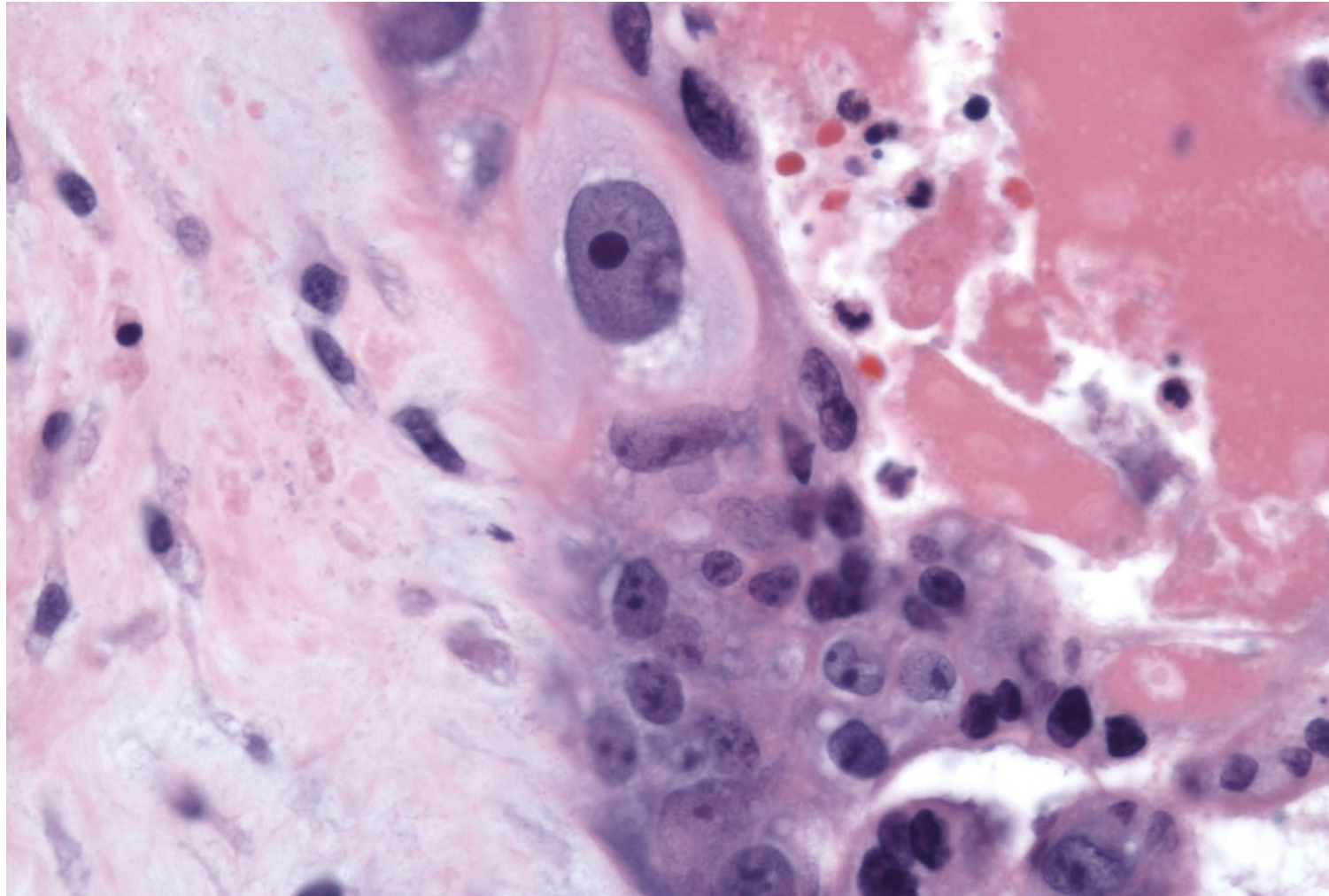
CHM are distinct from the normal progression from proliferation to differentiation



**MeIcAM | CD146**

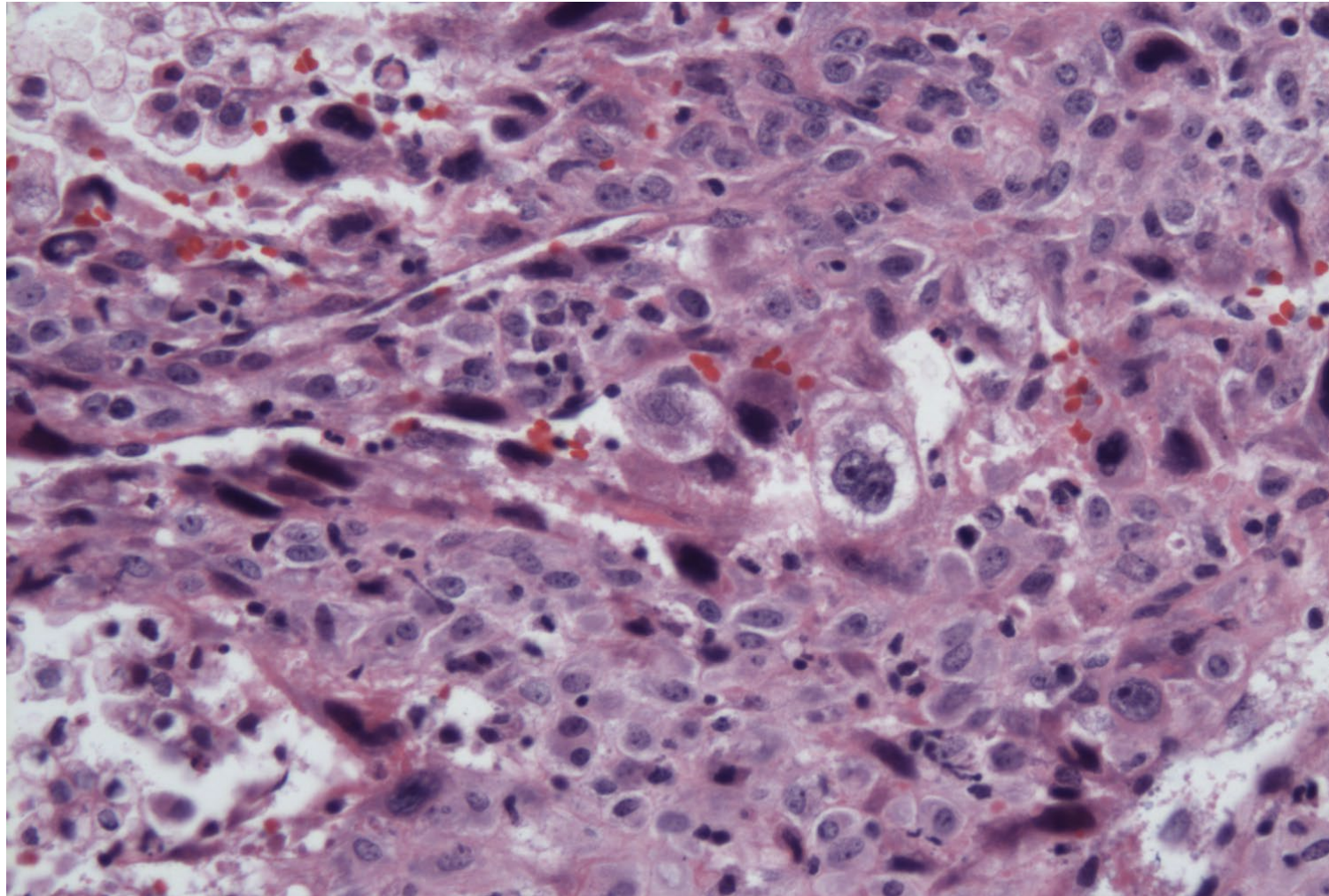
**Ki-67 | MIB1**





## Complete Hydatidiform Mole

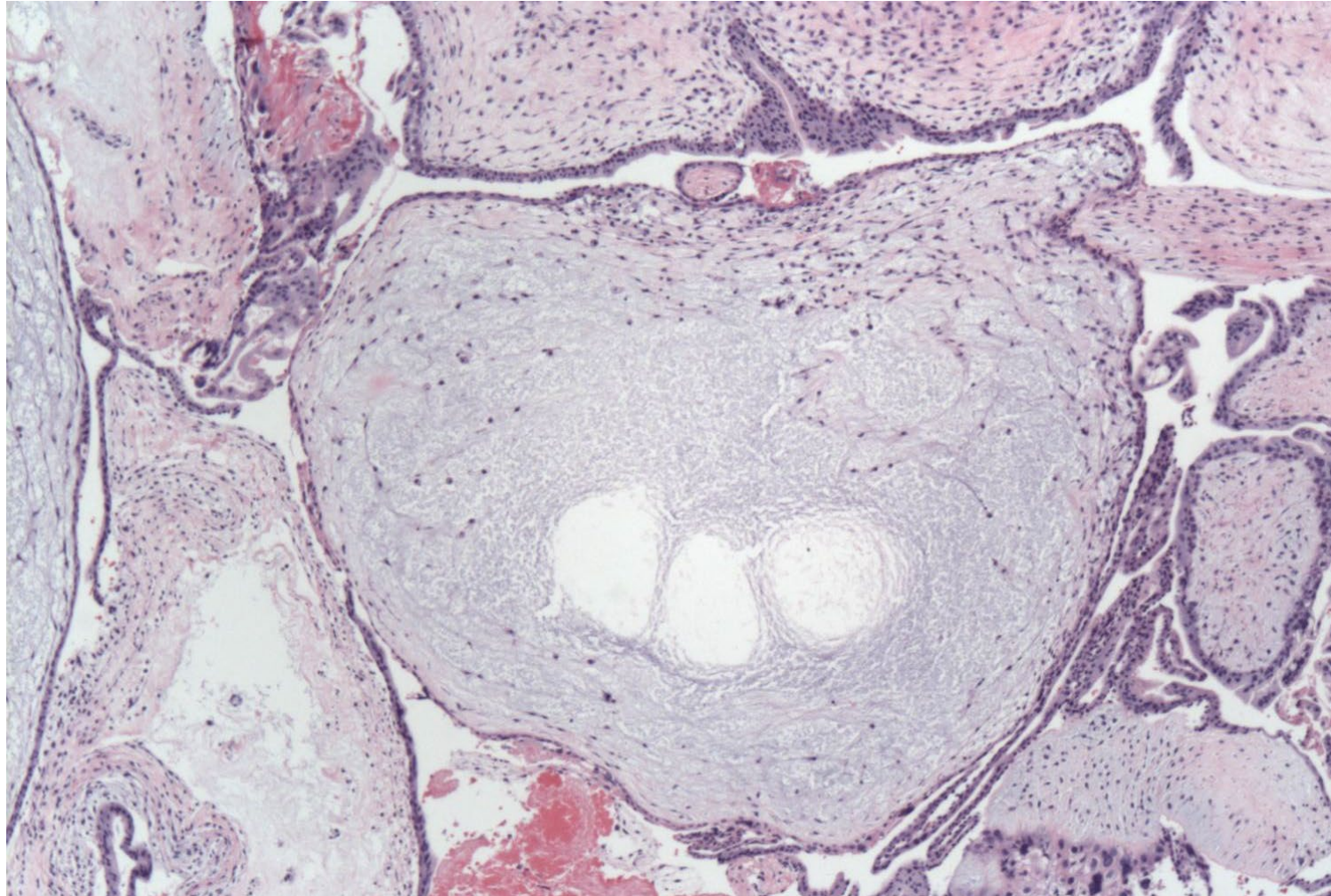
- **Trophoblast atypia**
  - **May range from modest to severe**



## Molar Implantation Site

- **Distinctive – easy to see; more atypical than usual**
- **May be your first clue**
- **Pitfall: mimics ChorioCa or PSTT**





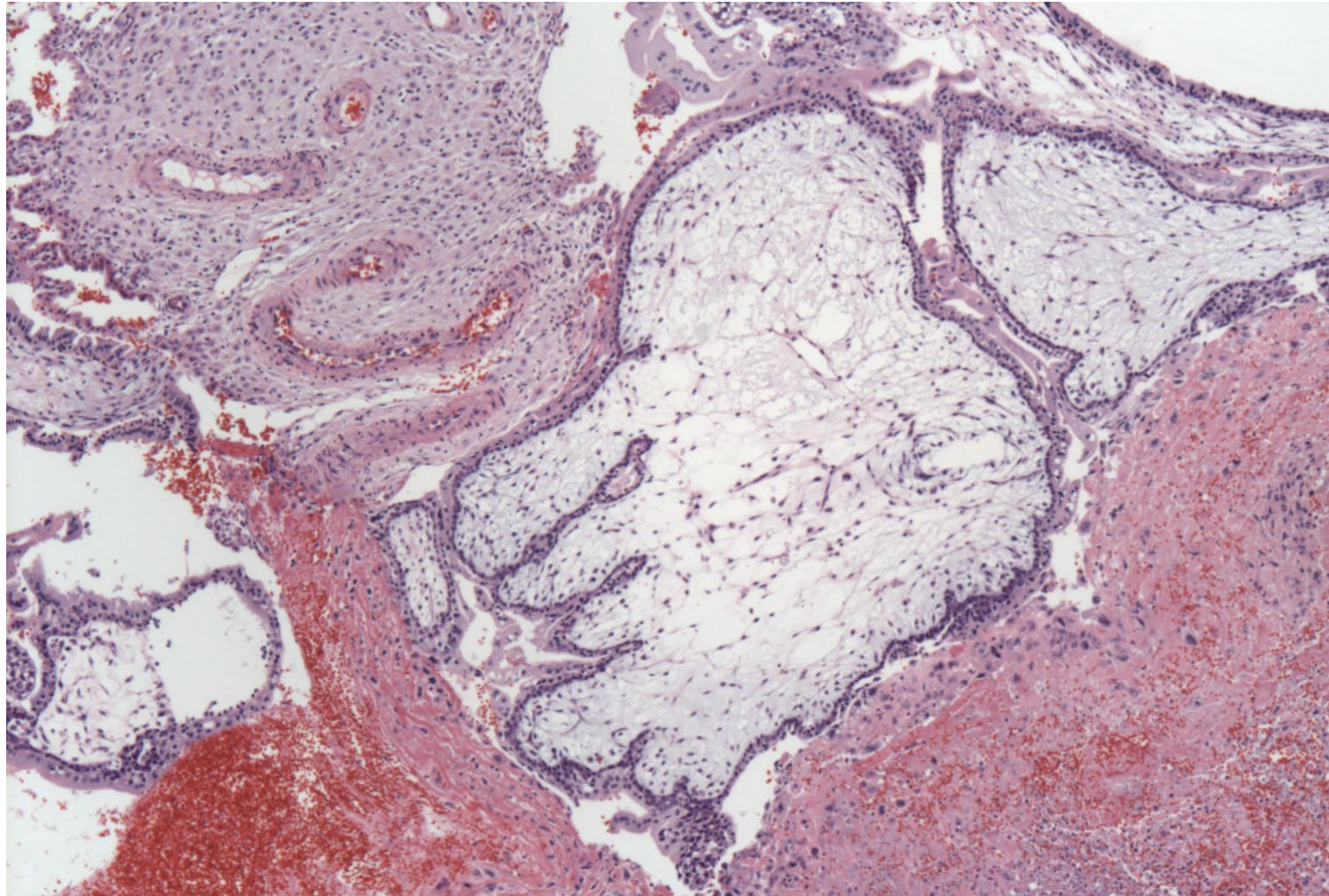
## Complete Hydatidiform Mole

**“Degenerating” CHM have less atypia and hyperplasia, may be confused with non-molar hydropic villi.**



# Early Complete Hydatidiform Mole





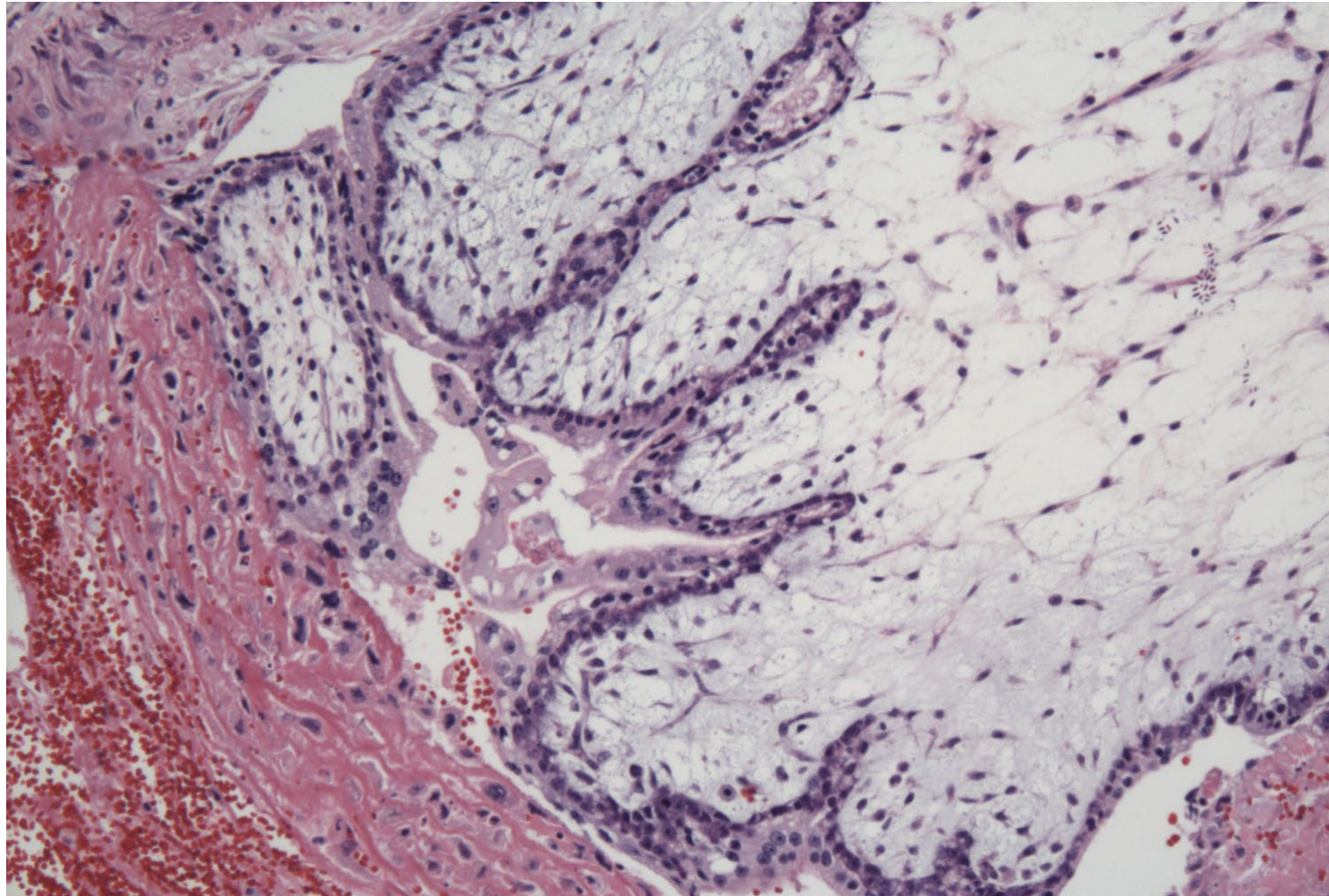
## Early Complete Mole

**Note:**

**Blue myxoid stroma**

**Pitfall: also found in very  
early gestations**



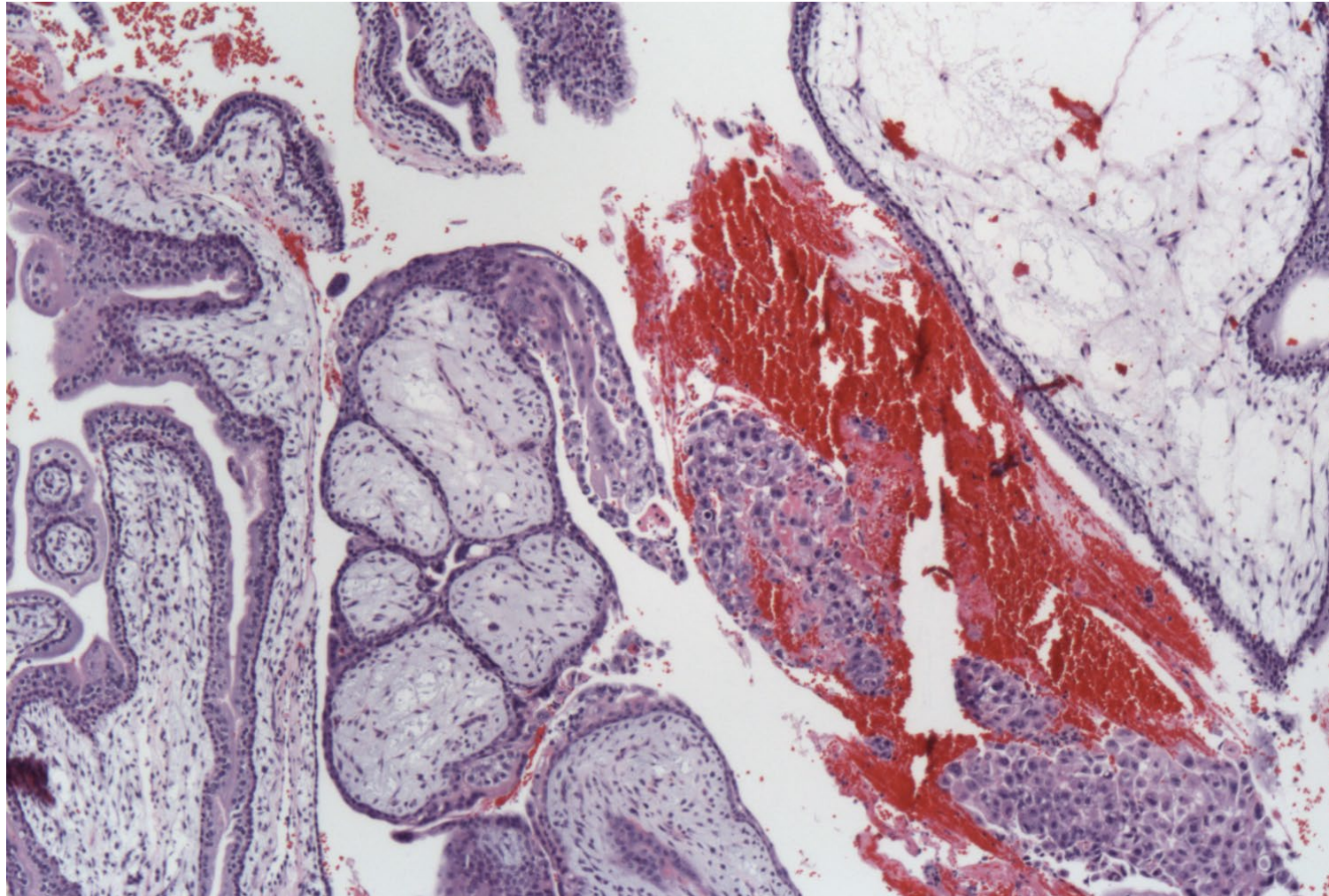


## Early Complete Mole

**Note:**

**Deep cleft-like infolding of  
trophoblast layer**



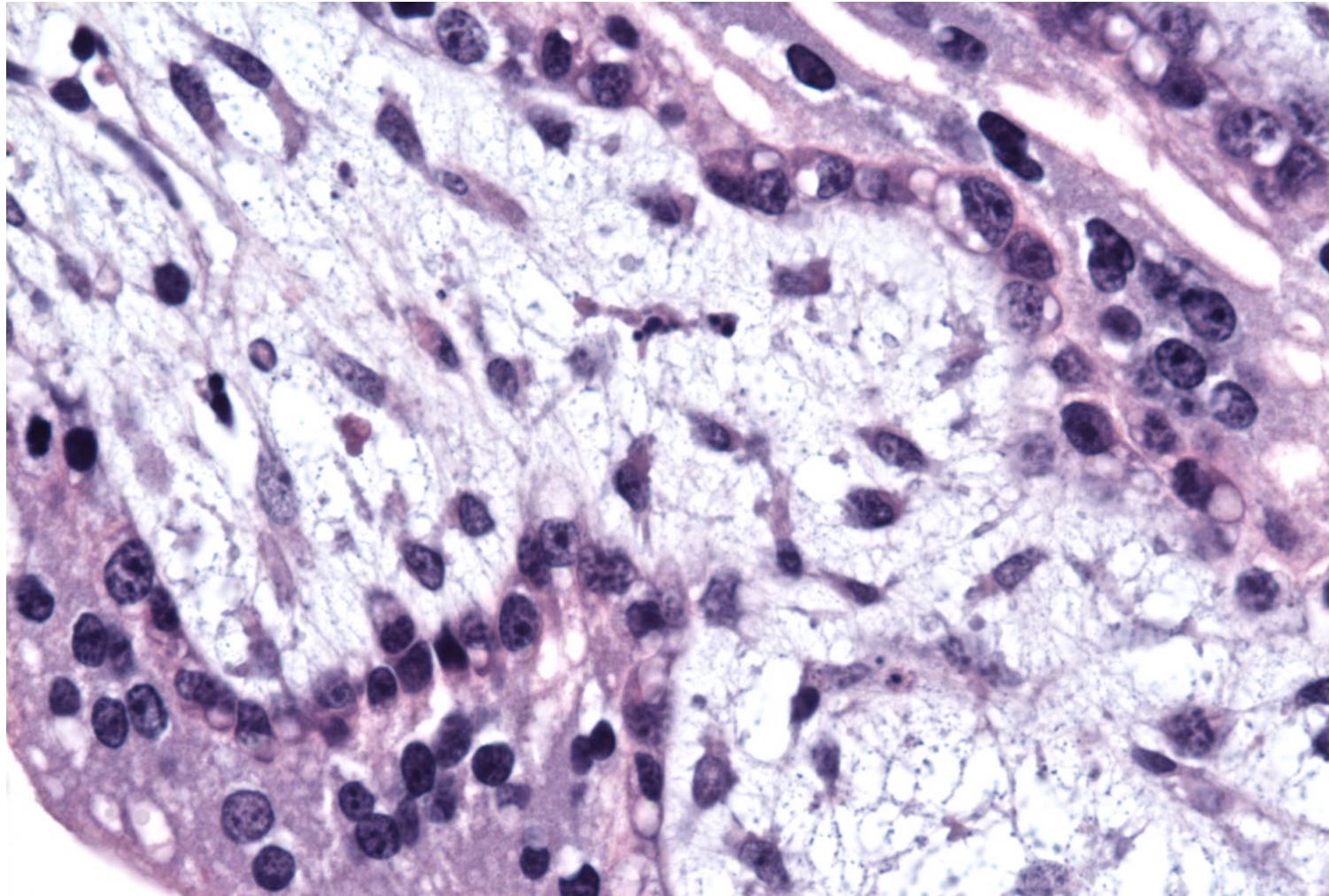


## Early Complete Mole

**Note:**

**Features of both Early and  
Classical CHM**



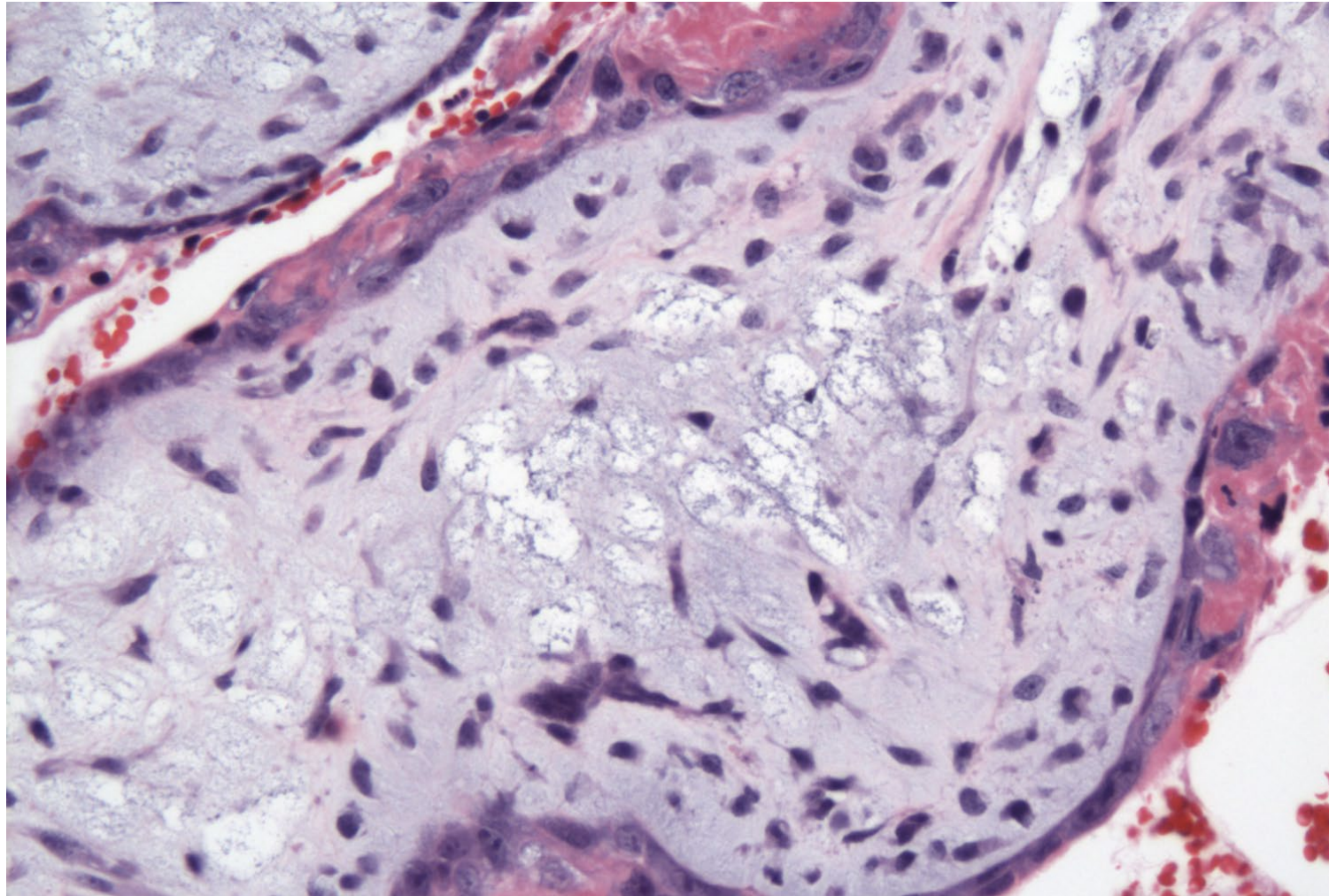


## Early Complete Mole

**Note:**

**Stromal (and vascular) cell  
degeneration**

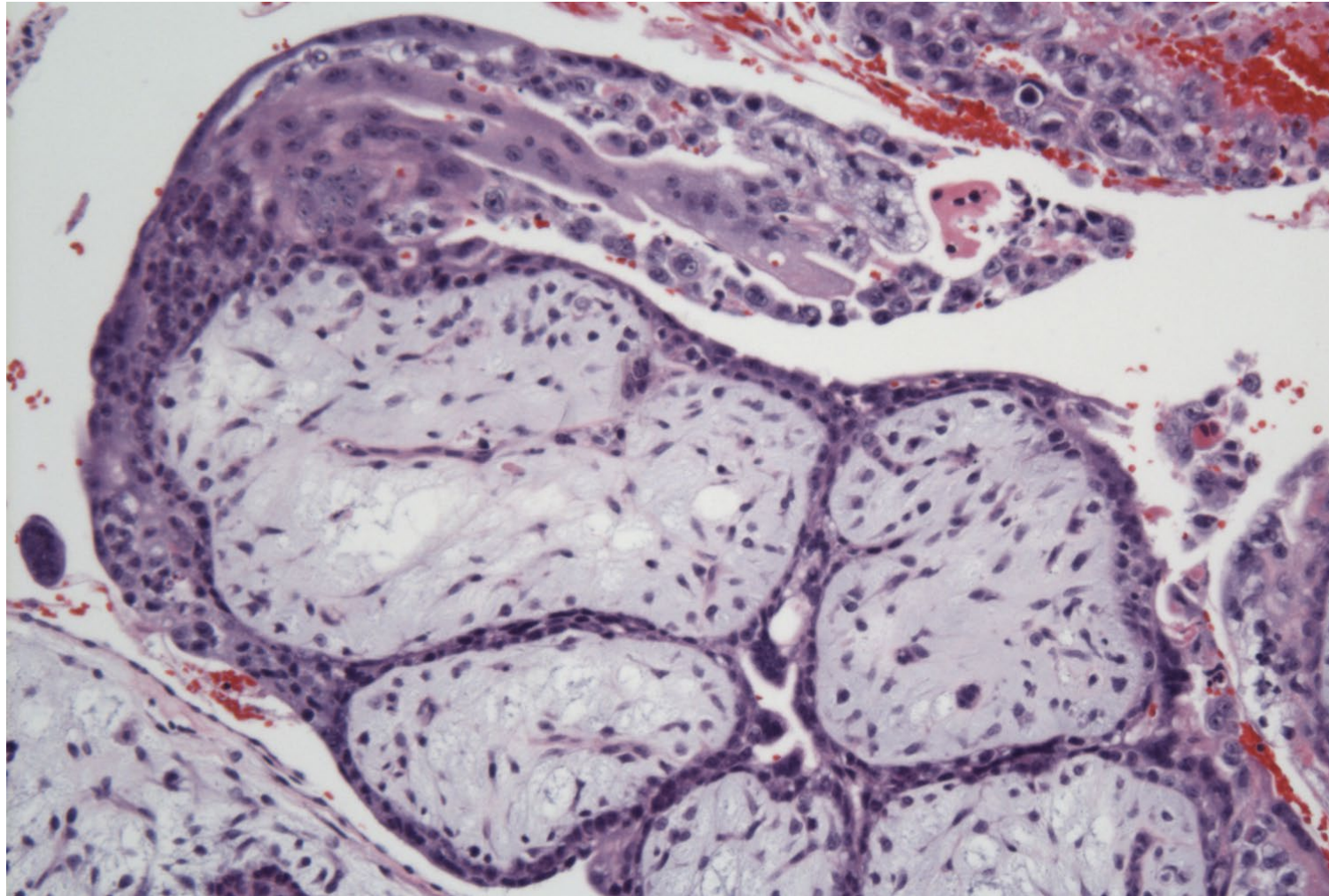




## Early Complete Mole

**Note:**

**Early cavitation**



## Early Complete Mole

**Note:**

**Early presentation more common intensively US screened populations**



# Complete Hydatidiform Mole

## Differential Diagnosis

- Very Early Gestational Sac
- Early Abortus
- Hydropic Abortus
- Partial Mole
- Twin Gestation with Complete Mole as Co-Twin

# Early Gestational Sac

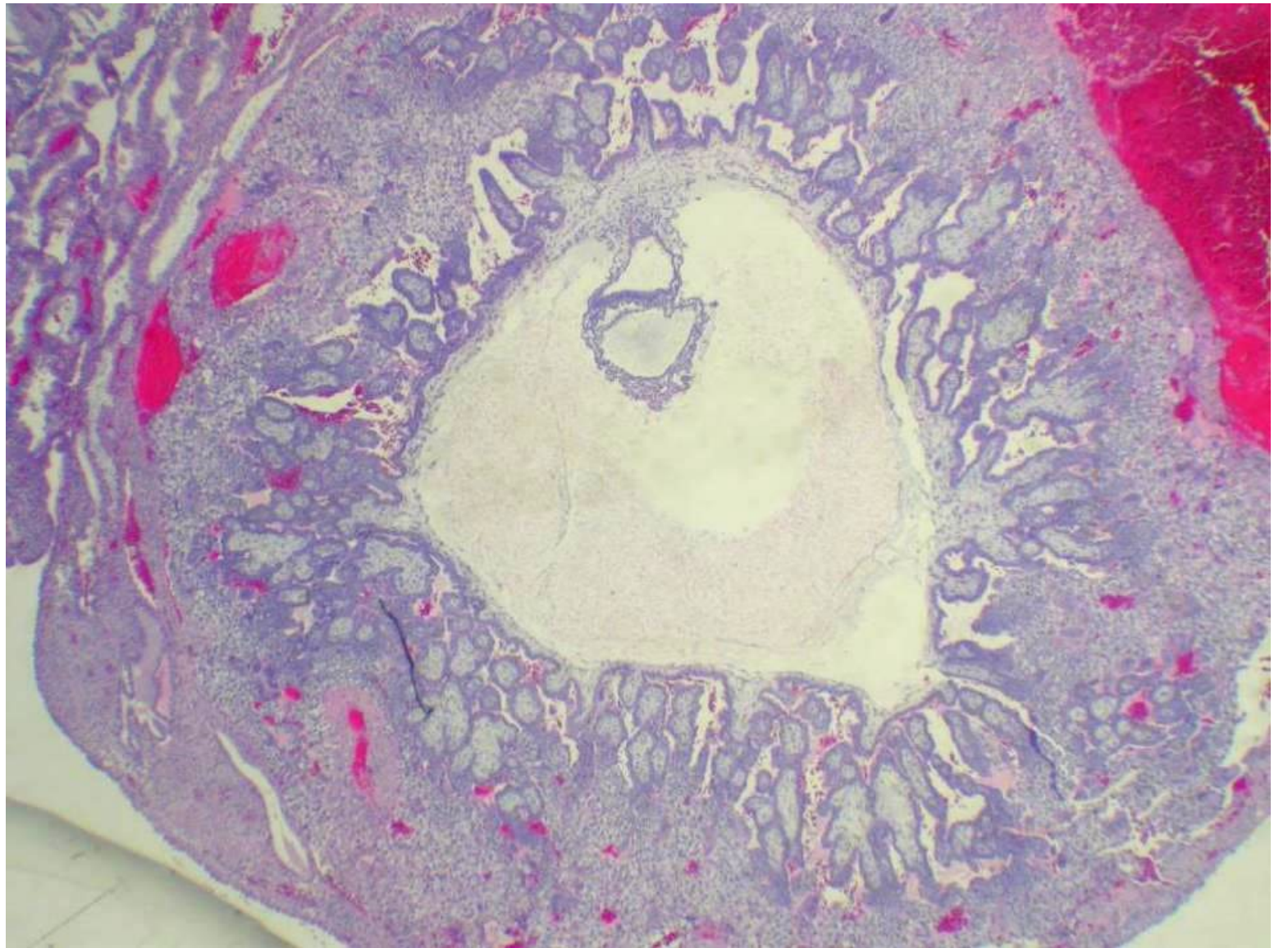
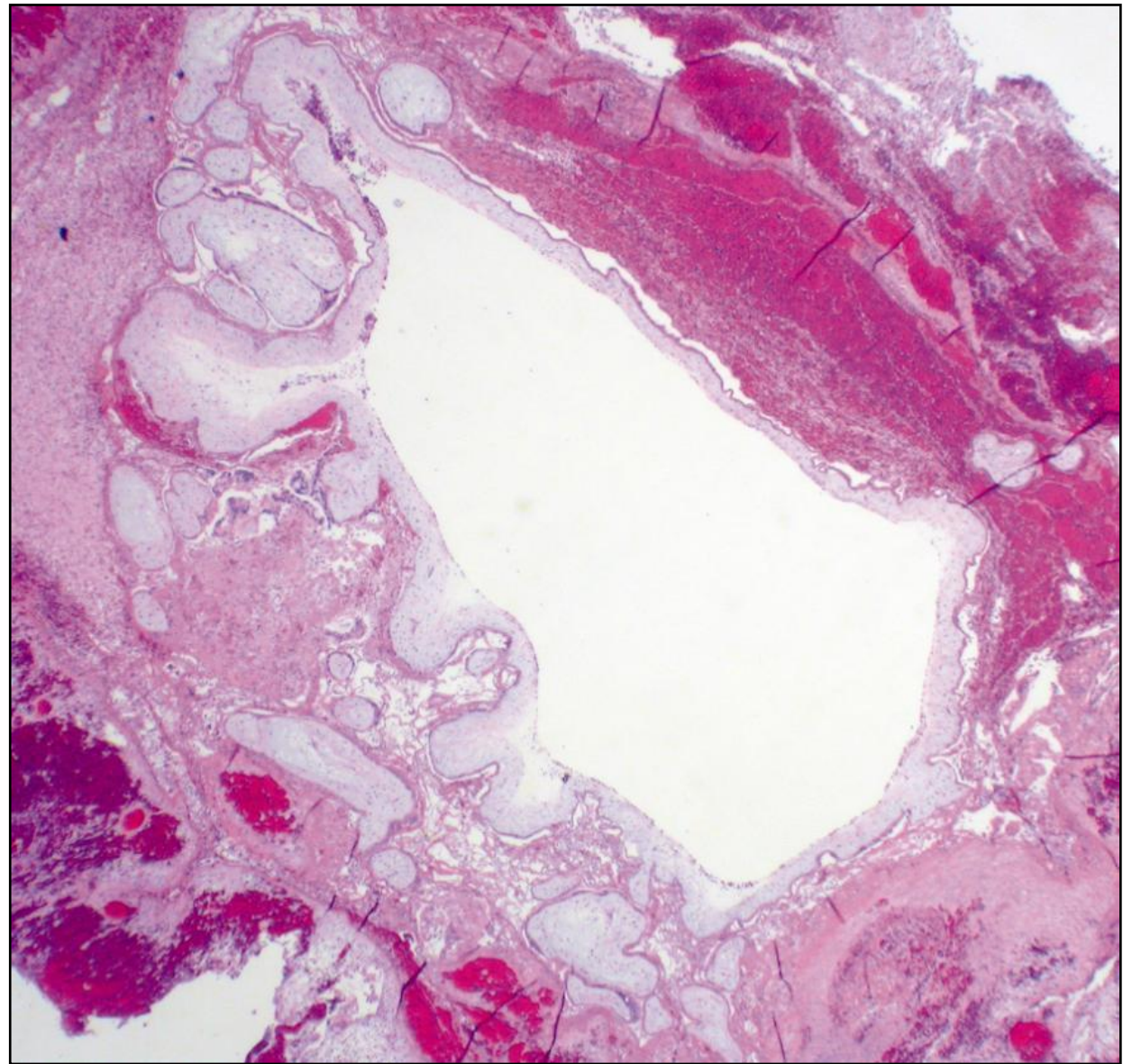


Image courtesy of Dr. T. Boyd



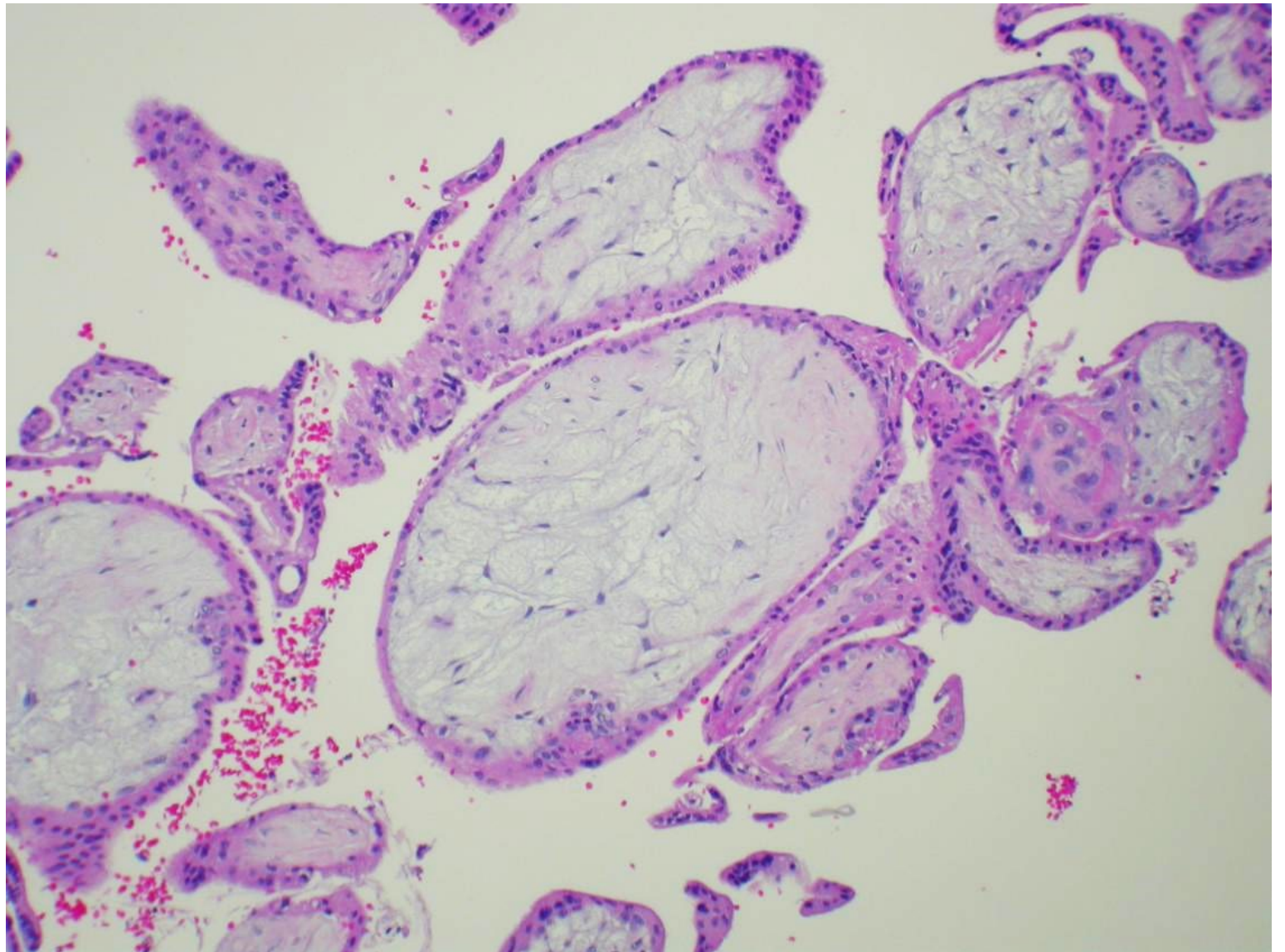
# Early Gestational Sac



Clue – Never more than a single cavitated villi

Image courtesy of Dr. T. Boyd

## Early Gestational Sac



Clue – Symmetric swelling with attenuation  
(no hyperplasia), without cavitation

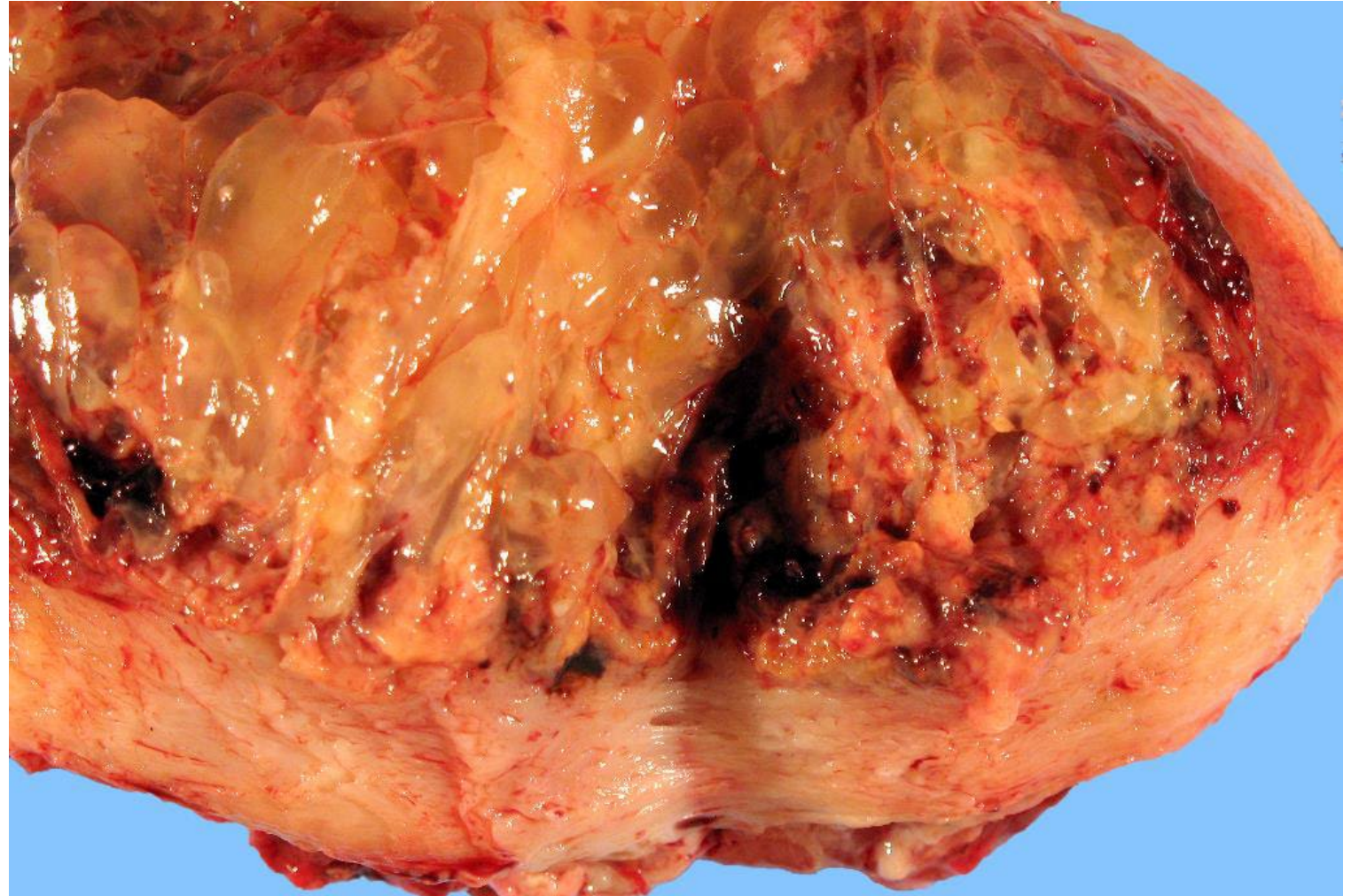
Image courtesy of Dr. T. Boyd



Invasive  
Complete Mole

“Chorioadenoma  
Destruens”

CHM  
x  
Placenta Increta



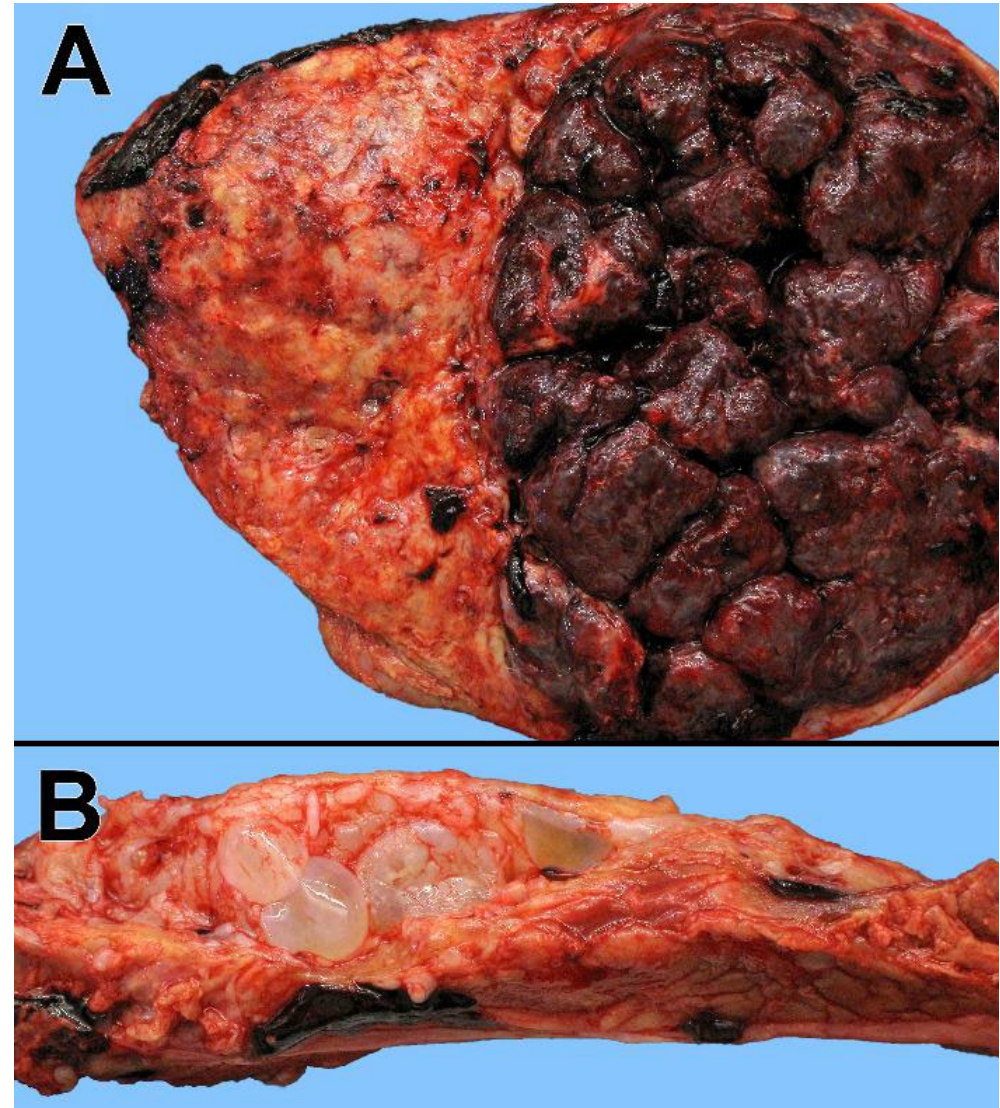
# Things to keep in mind about Invasive Mole

- Not readily detectible by curettage
- More frequent in age > 40 yr and underdeveloped countries
- Hemorrhage may obscure villi grossly
- Vascular space invasion common
- Frequency of persistent or metastatic gestational trophoblastic disease (GTD):
  - CHM >> *Invasive Mole* > ChorioCA
- Villi may be scarce
- Pathobiologic mechanism unknown



# Complete Hydatidiform Mole

in a  
Twin Gestation



# Things to keep in mind about Molar Twins

- Rare
- CHM of interest, but PHM also possible as co-twin
- Pitfall – iatrogenic admixture of CHM co-twin by curettage mimics PHM
- Presence of normal co-twin often delays diagnosis
- Higher pre-evacuation  $\beta$ -hCG; clinical symptoms more likely
- Poses obstetrical management issues, but successful deliver possible



# Ancillary tools for complete mole

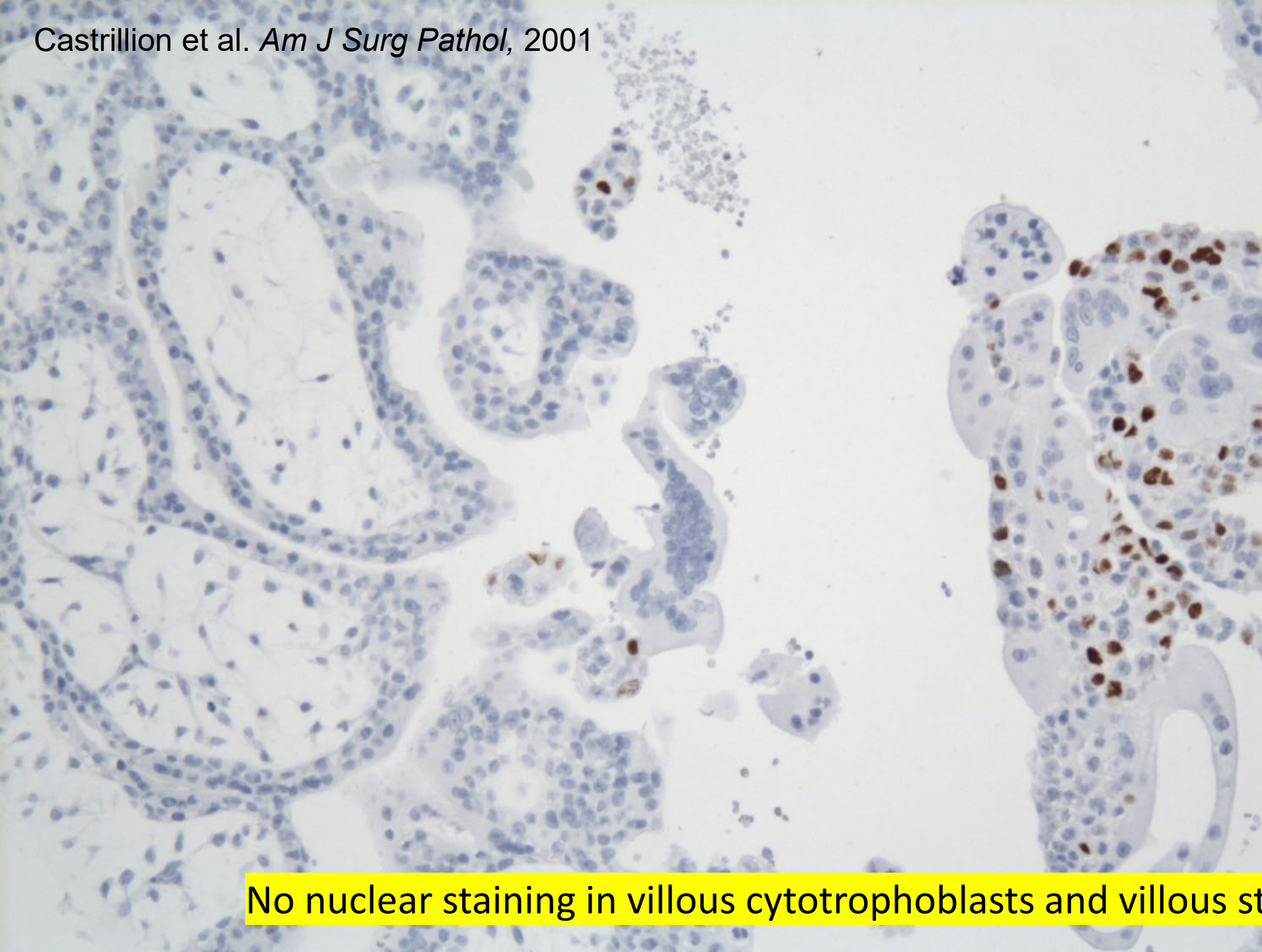
- Genetics
  - Flow cytometry for DNA index ( 2n or 4n ploidy)
  - Karyotype (46,XX or 46,XY)
  - Microarray
    - SNP analysis may detect genome-wide LOH (uniparental) if derived from single sperm
  - Microsatellite polymorphisms
    - Diandry if parental DNAs collected
- Immunohistochemistry
  - Leverage underlying pathobiology of aberrant imprinting

# cyclin-dependent kinase inhibitor 1C (p57, Kip2) (CDKN1C)

- Imprinted, with preferential expression of the maternal allele
- Strong inhibitor of several G1 cyclin/Cdk complexes and a negative regulator of cell proliferation
- Tumor suppressor candidate and plays a role in Beckwith-Wiedemann syndrome
- Part of an imprinted gene region on 11p15.5 with IGF2 and H19



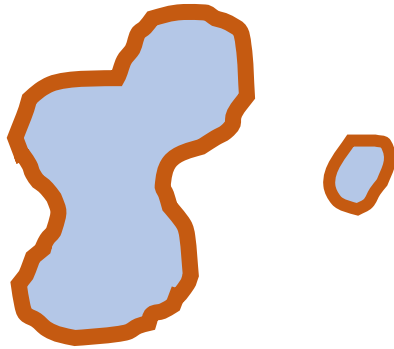




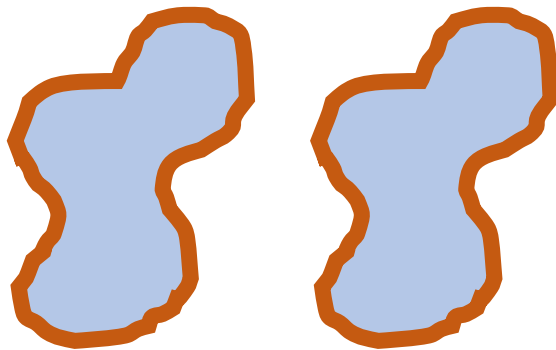
Relaxation Of Imprinting  
In  
Extra-Villous Trophoblasts

No nuclear staining in villous cytotrophoblasts and villous stromal cells

# Murphy's Law for Reproduction – Watch out for unusual p57 staining patterns!!



Twin with CHM Co-Twin



Chimeric/Mosaic Gestation

Some behave like CHM

Also seen in PHM mimics

Placental Mesenchymal Dysplasia

Beckwith–Wiedemann syndrome



# Partial Hydatidiform Moles



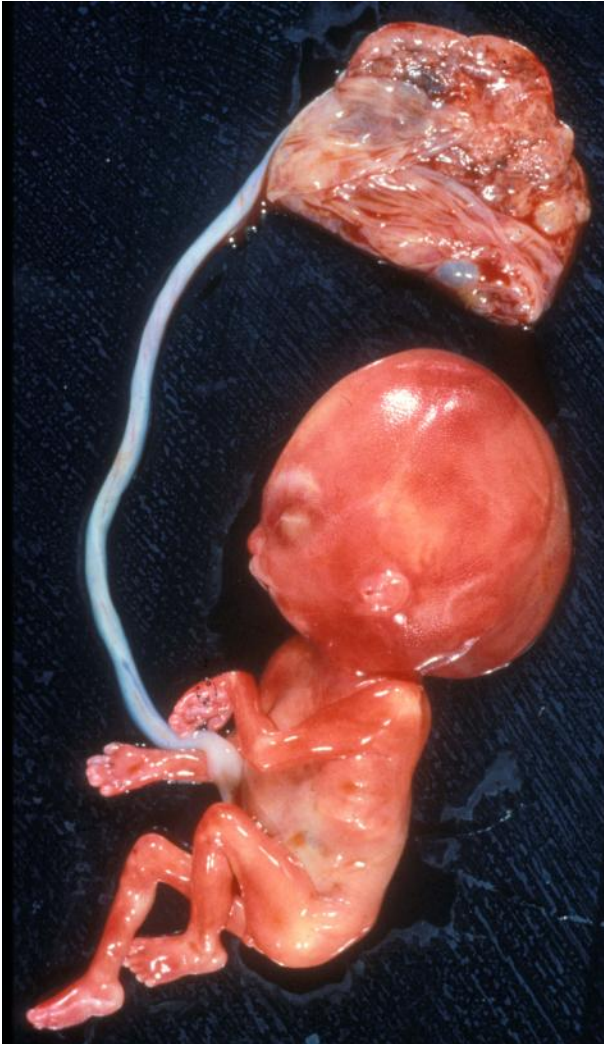


## Partial Hydatidiform Mole

- Image courtesy of Dr. T. Boyd



# Partial Hydatidiform Mole – Maldeveloped fetuses with 3-4 syndactyly

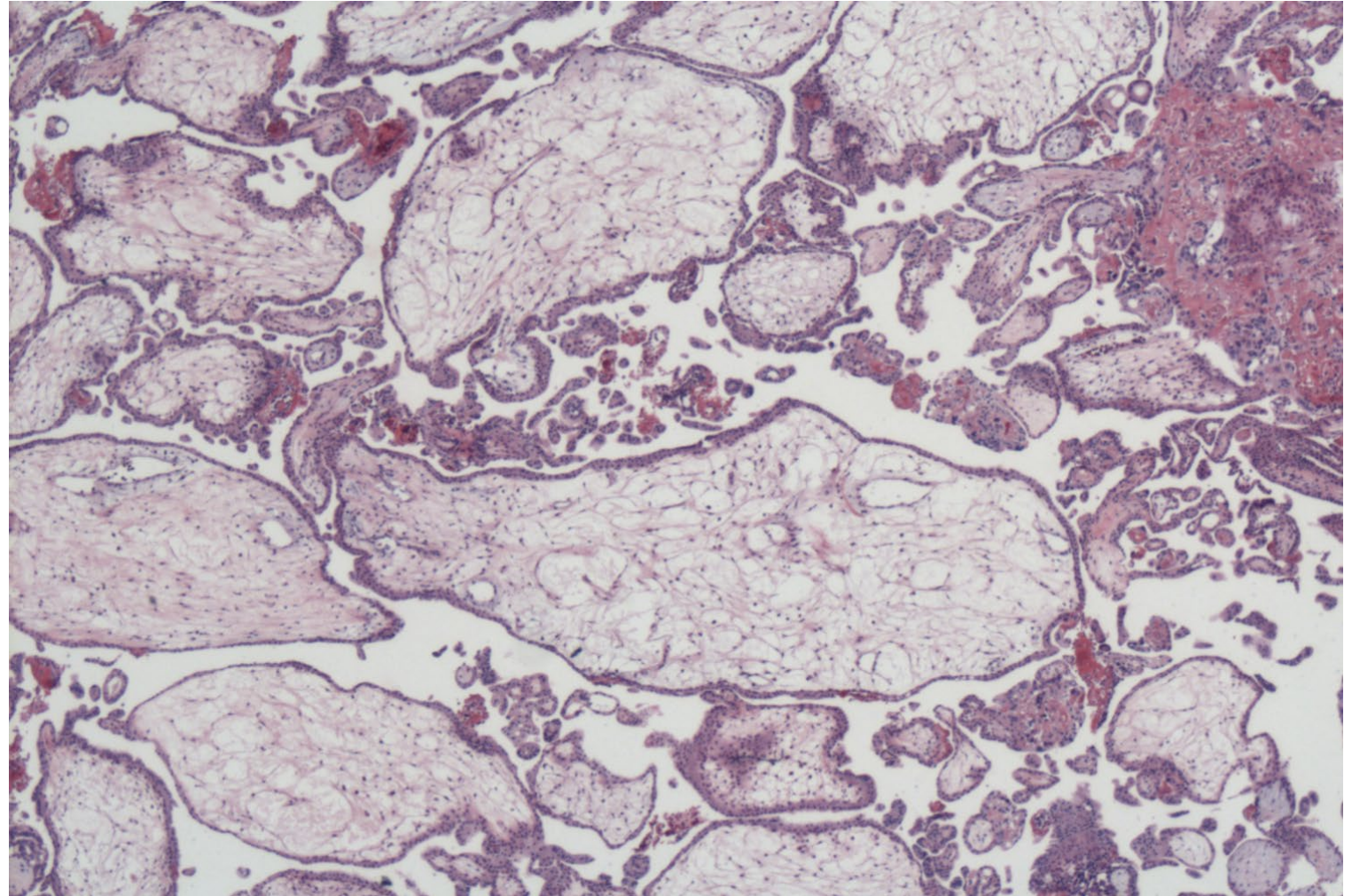


Images courtesy of Dr. T. Boyd

# Partial Hydatidiform Mole

Intermixed  
Biphasic  
Population of  
Villi by Size

proposed stringent  
threshold for villous  
enlargement:  
>2.5 mm



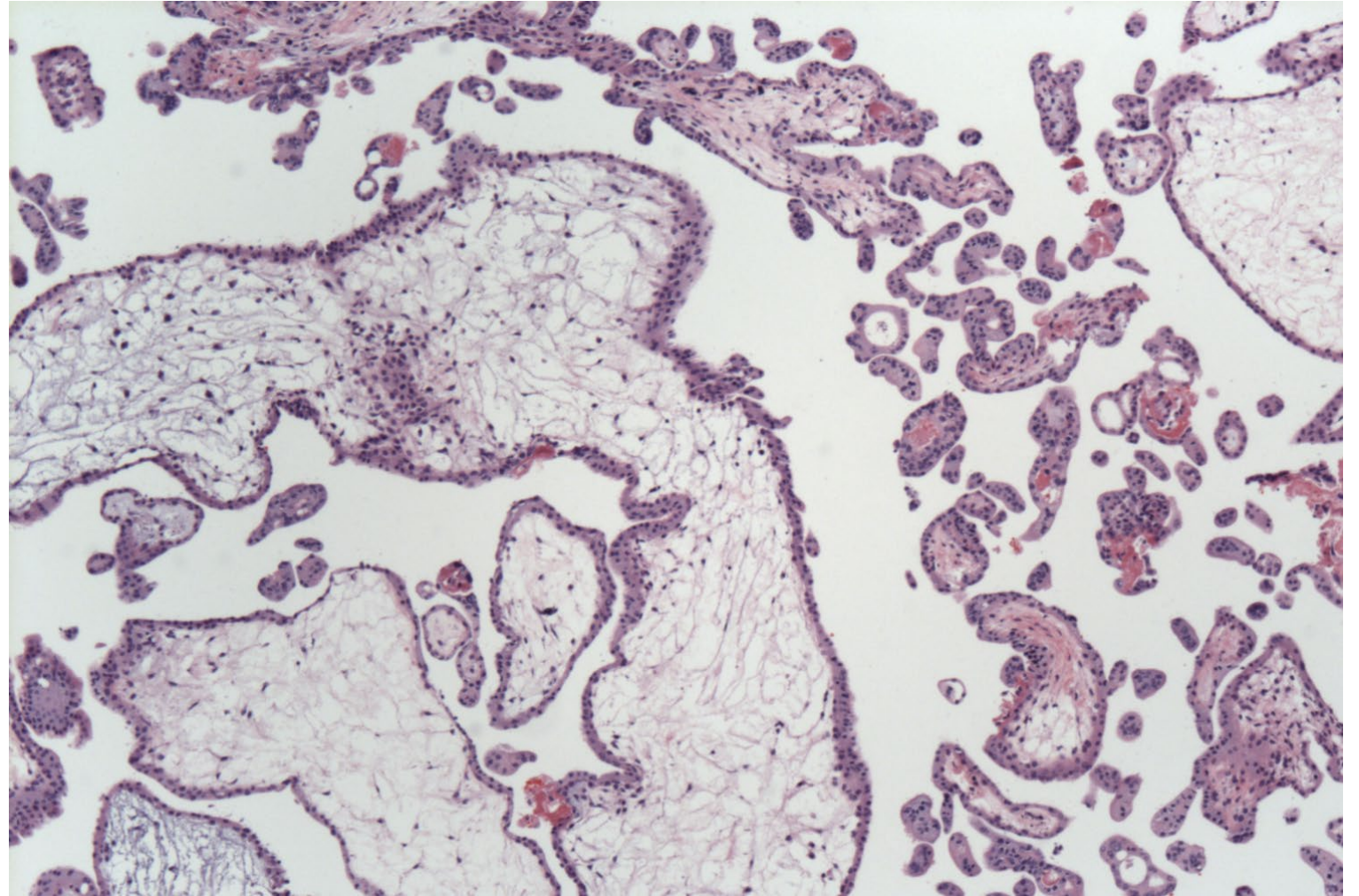


# Partial Hydatidiform Mole Have Irregularly Shaped Enlarged Villi

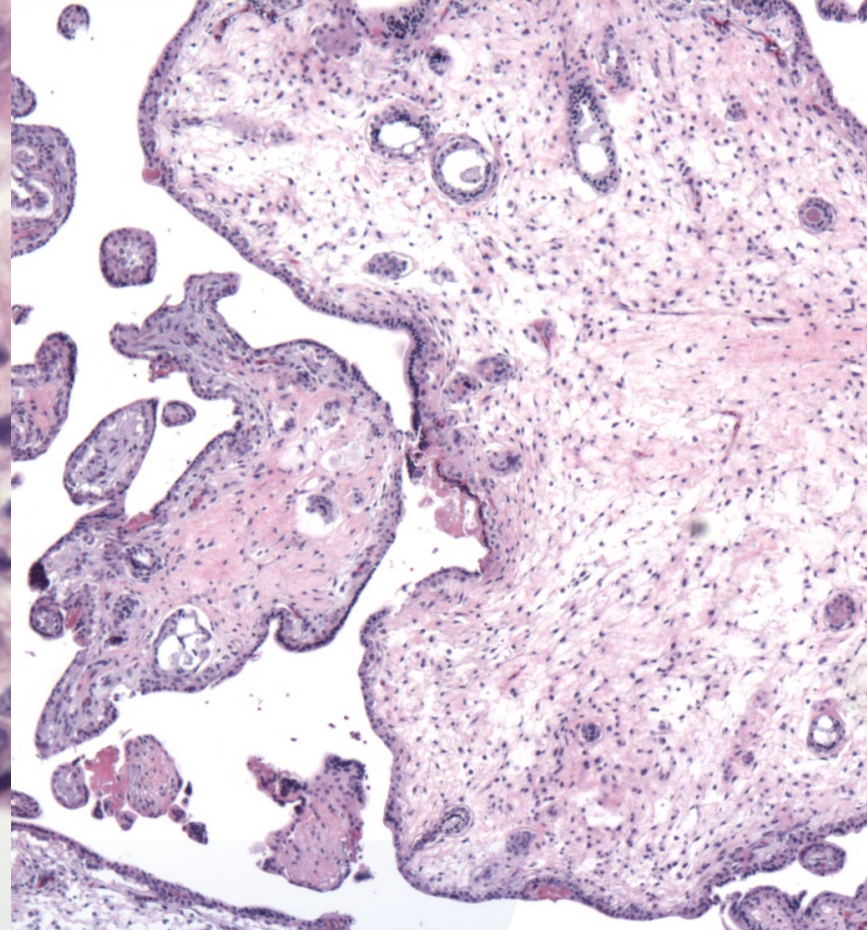
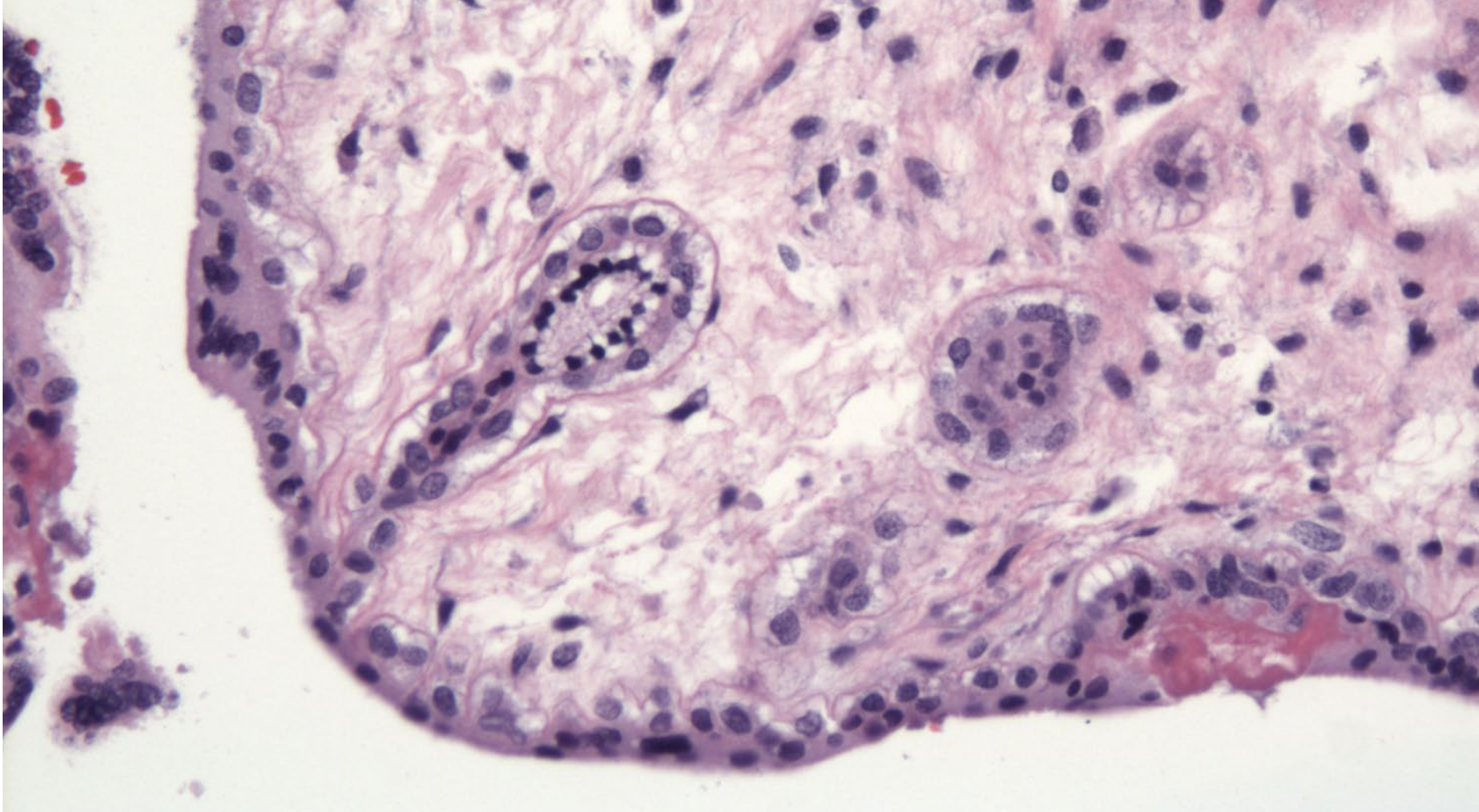


# Partial Hydatidiform Mole

Irregular  
Shapes in  
Larger Villi







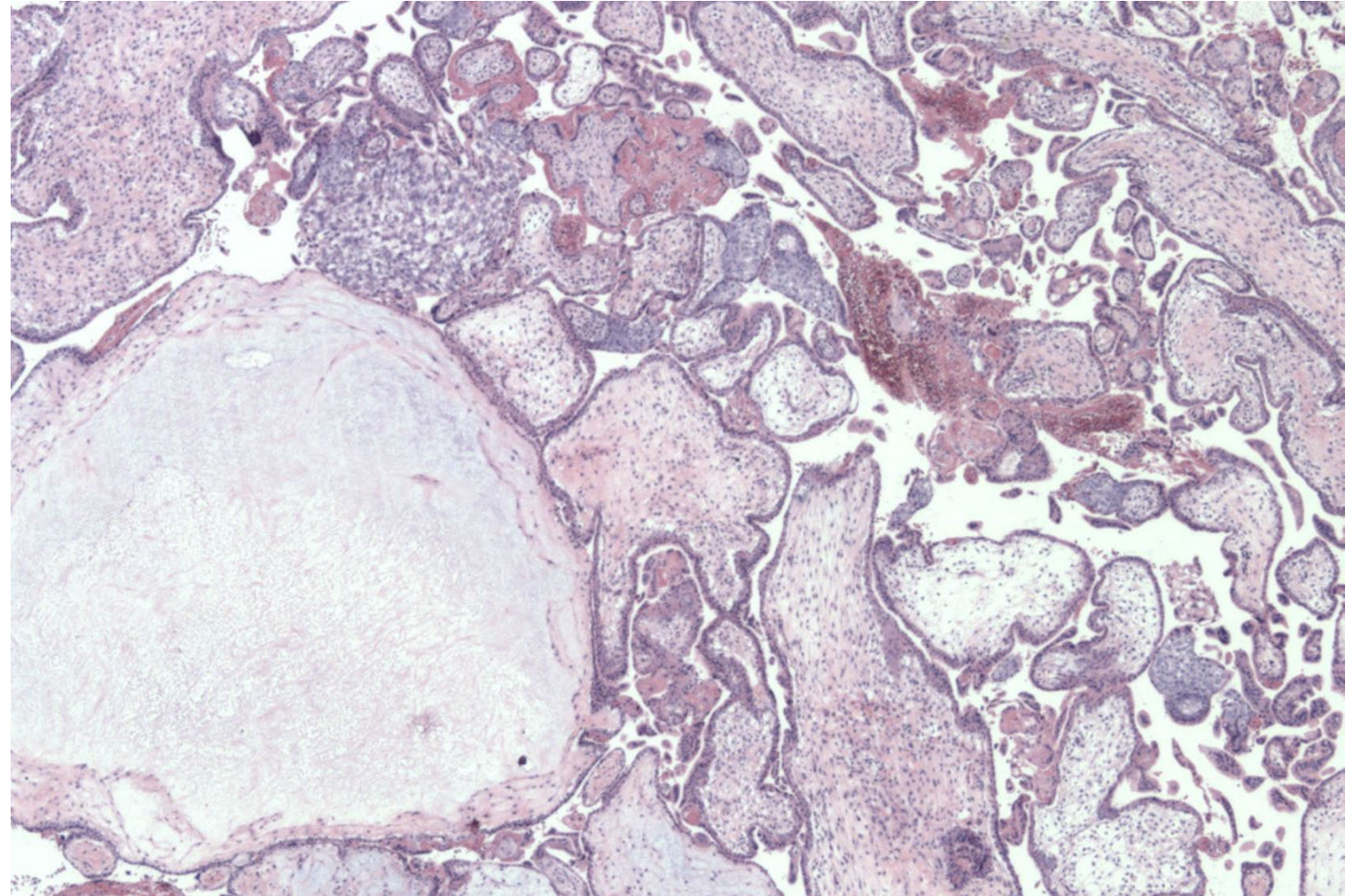
Partial Hydatidiform Mole –  
Inclusions Reflect Shape Complexity



# Partial Hydatidiform Mole

Scattered  
Cavitation of  
Larger Villi

my favorite criteria

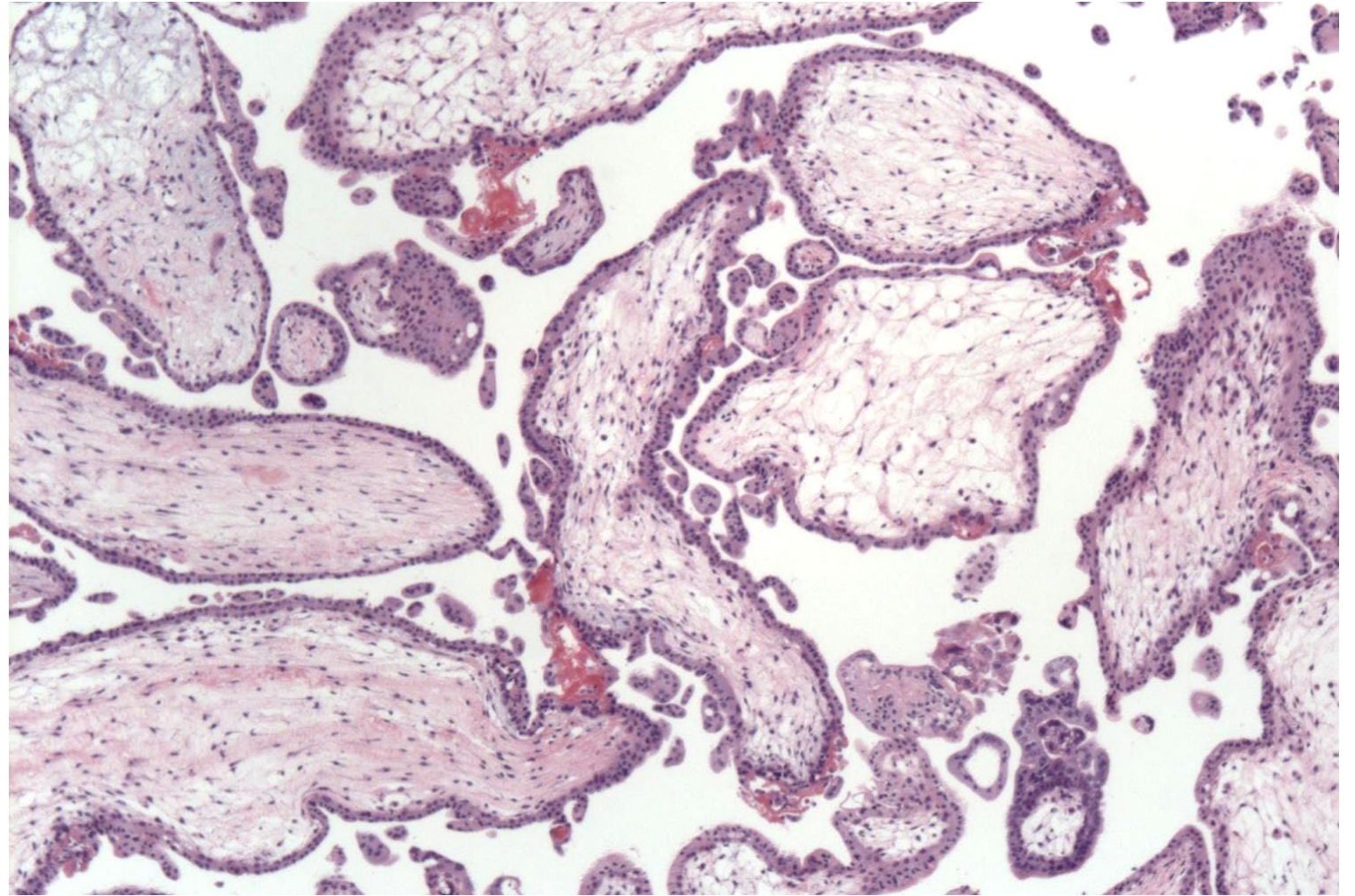




# Partial Hydatidiform Mole

## Trophoblastic Hyperplasia

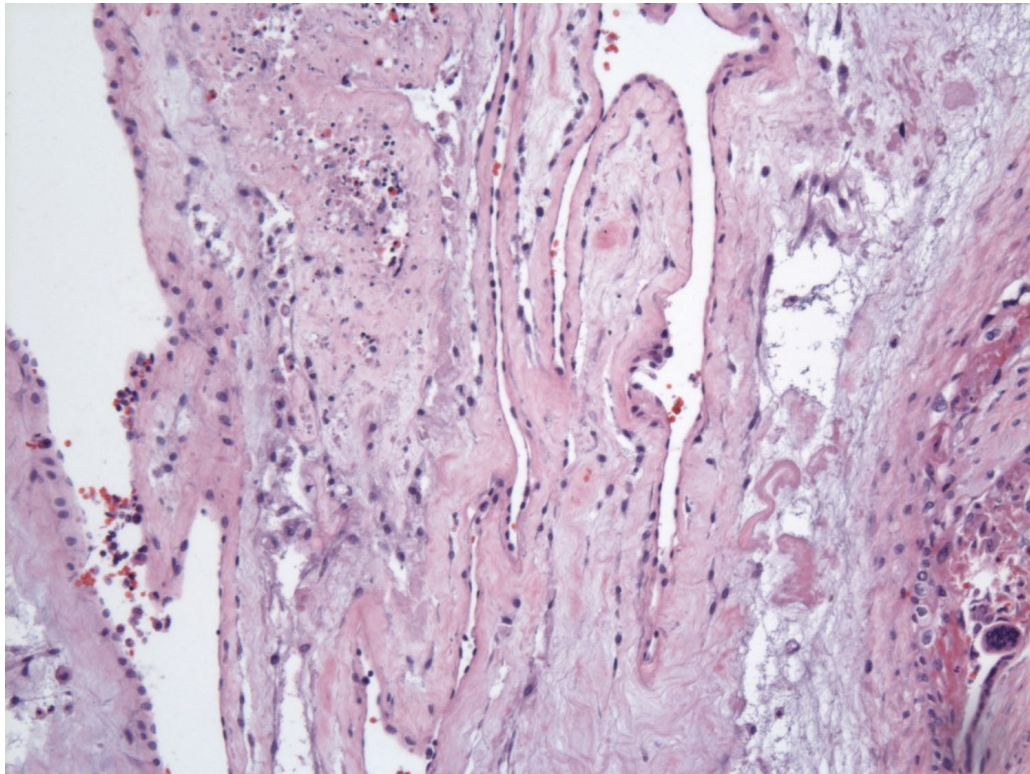
Proliferation  
commensurate to  
size & “tagging”



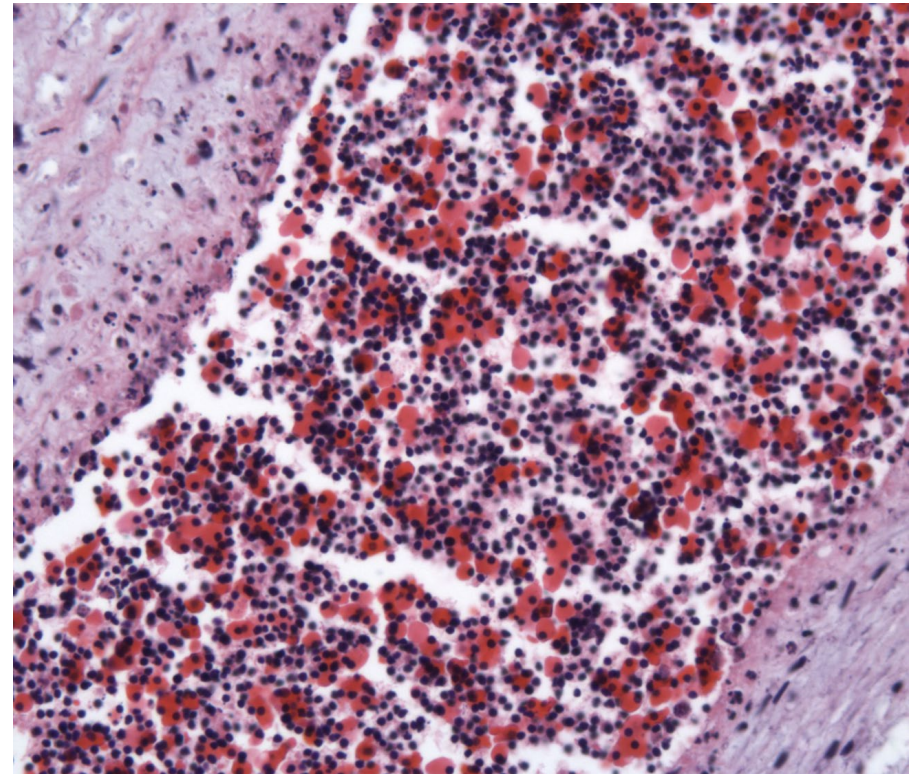


# Fetal Development in Partial Mole

**Amnion and Chorion**



**Fetal blood (common) and Embryo (uncommon)**

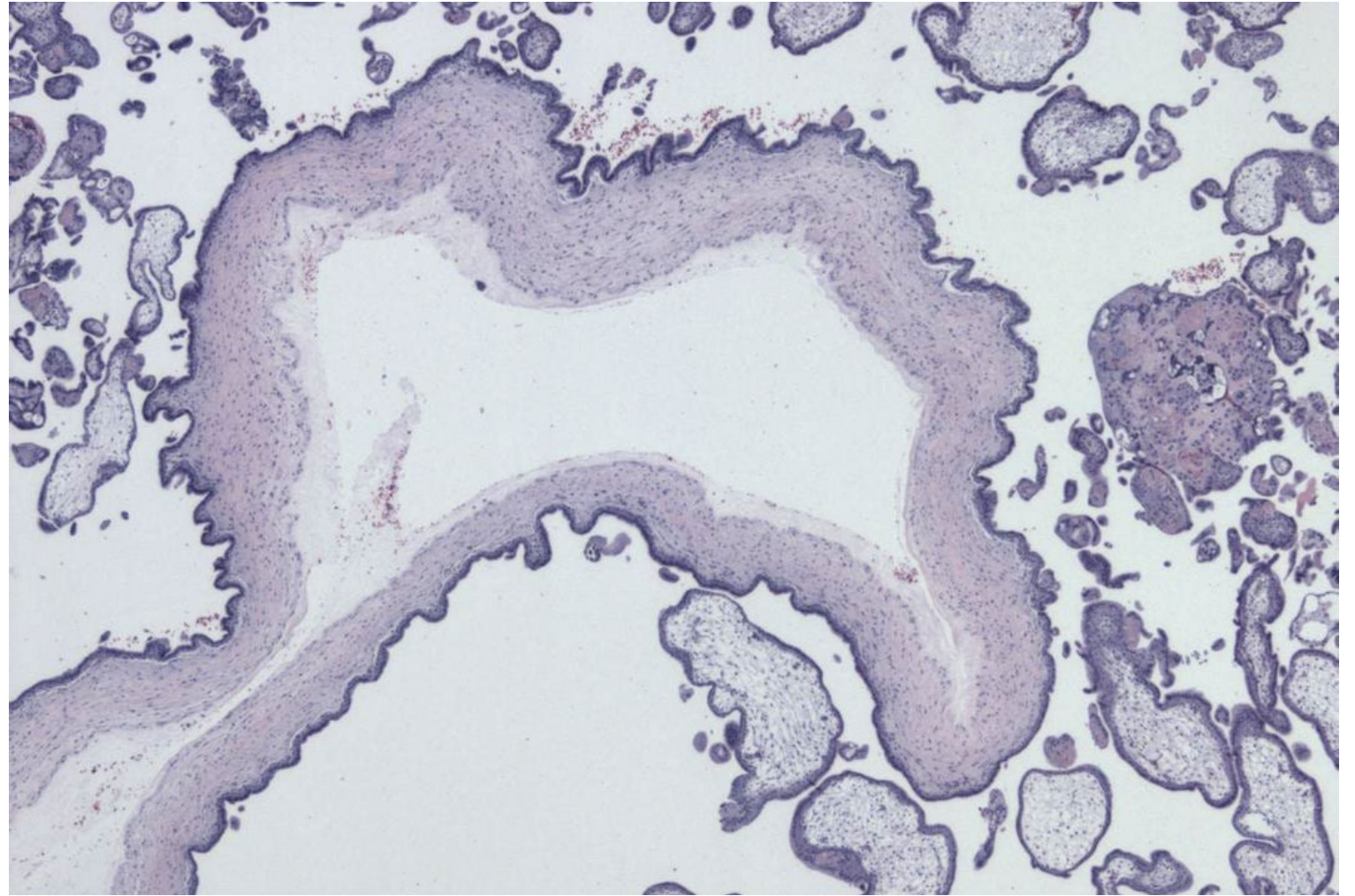


**Numerous Nucleated Fetal Red Blood Cells  
Not Typically Found In CHM**



Diploid partial  
mole mimic  
(ish CEP17x2,  
nl p57 IHC)

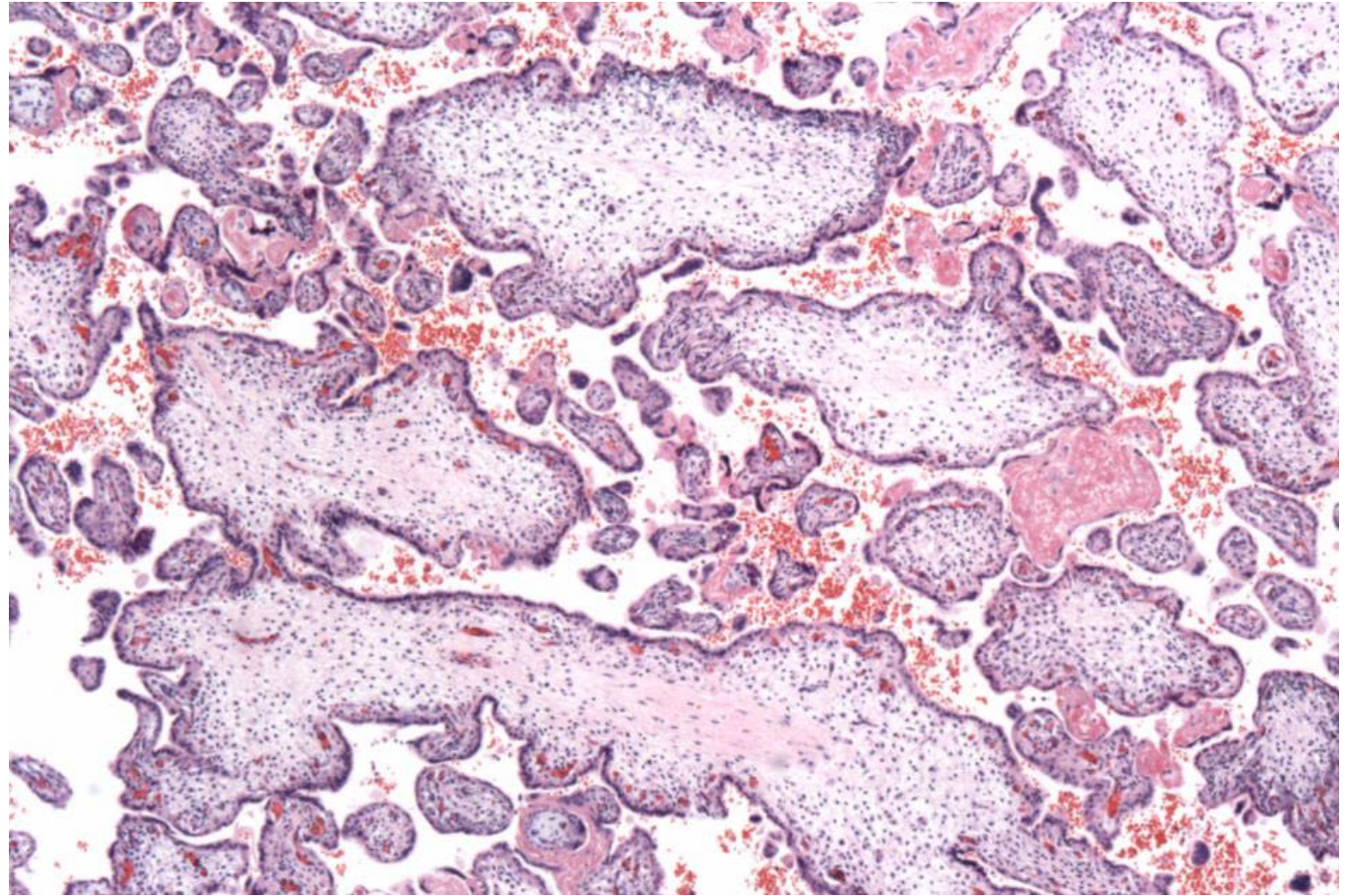
Chorionic  
plate  
mimicking a  
cavitated villus





Trisomy 21  
mimicking  
partial mole

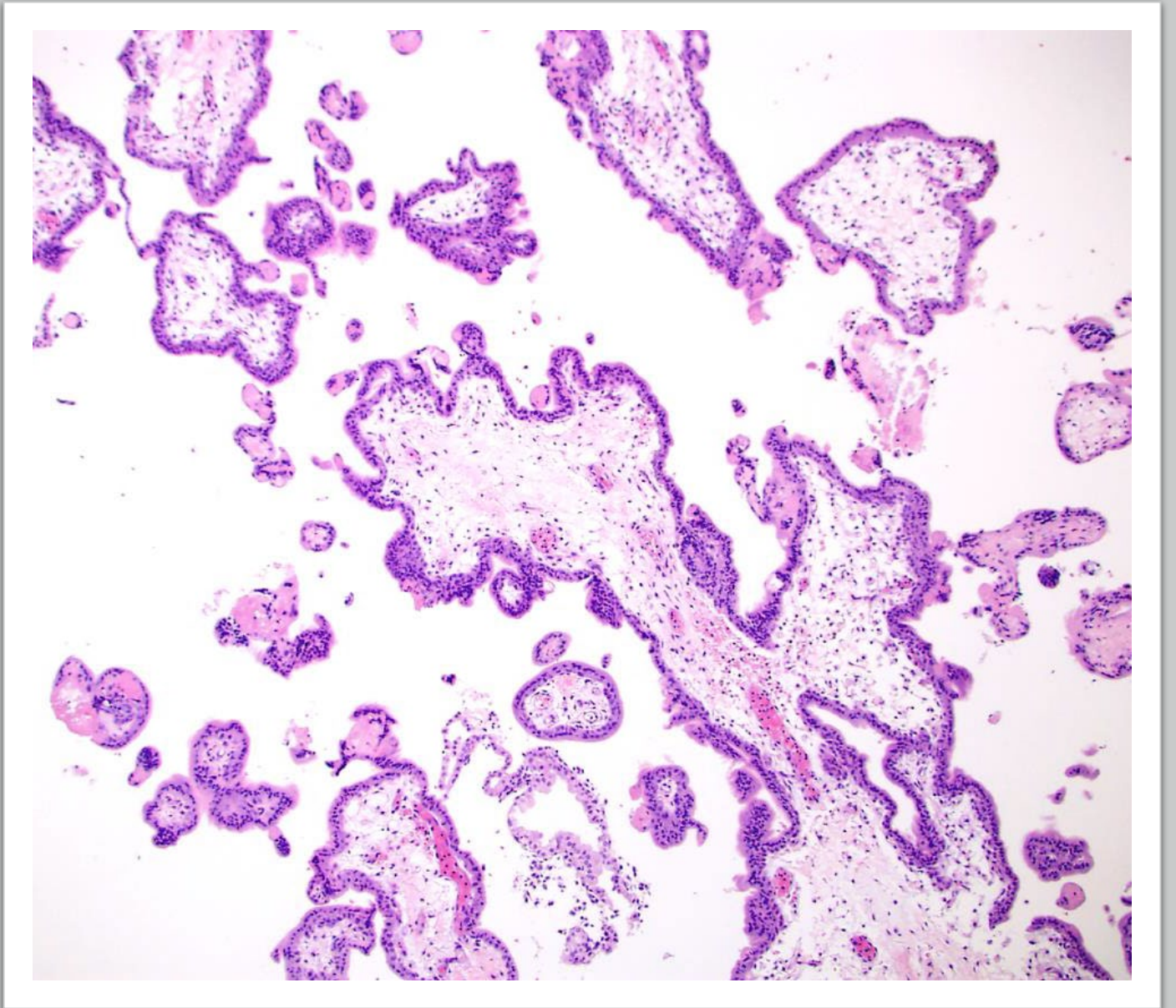
Dysmorphic  
villi with  
dimorphism  
but without  
cavitation





# Trisomy 13 is a very frequent PHM mimic

Best aneusomy mimic that I've seen was trisomy 11 (because it has 3 copies of p57 locus, presumable 2 from the father)



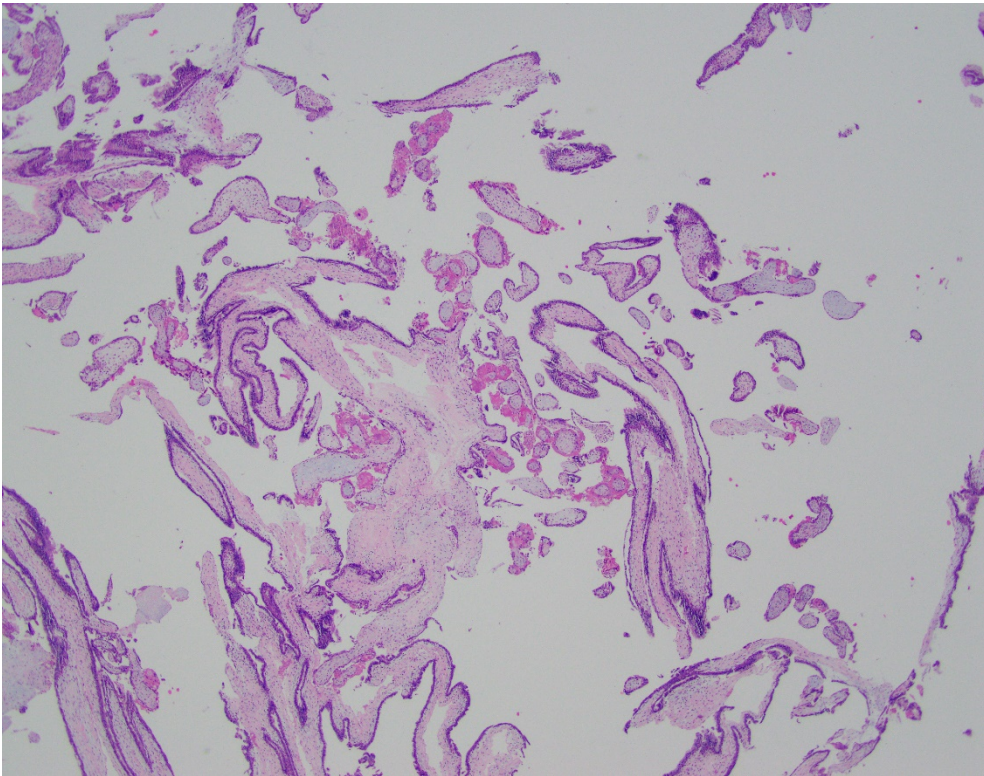
## Partial Hydatidiform Mole Are Diandric Triploid Gestations Favoring Placental Growth

	Redline et al. <i>Hum Pathol</i> , 1998	Han et al. <i>Am J Surg Pathol</i> , 2020	Various Case Reports
Starting population	Spontaneous abortion (832)	Prenatal screening for aneuploidy	
Refined population	Triploid (65)	Triploid (20)	
Methodology	Karyotype & Microsatellite PCR	Genotype Testing	
Digynic (1P:2M)	Non-molar (31%)  Fetal tissue more frequently found	Non-molar (35%)	
Diandric (2P:1M)	Partial mole (61%)  phenotypic spectrum including “early”, “ancient”, “suggestive”	Partial mole (65%)  Only ¼ diagnostic by (+) cistern and >2.5 mm  ¾ had focal or incomplete phenotype	
Tetraploid Triandric (3P:1M)			Rare cases with features resembling partial mole

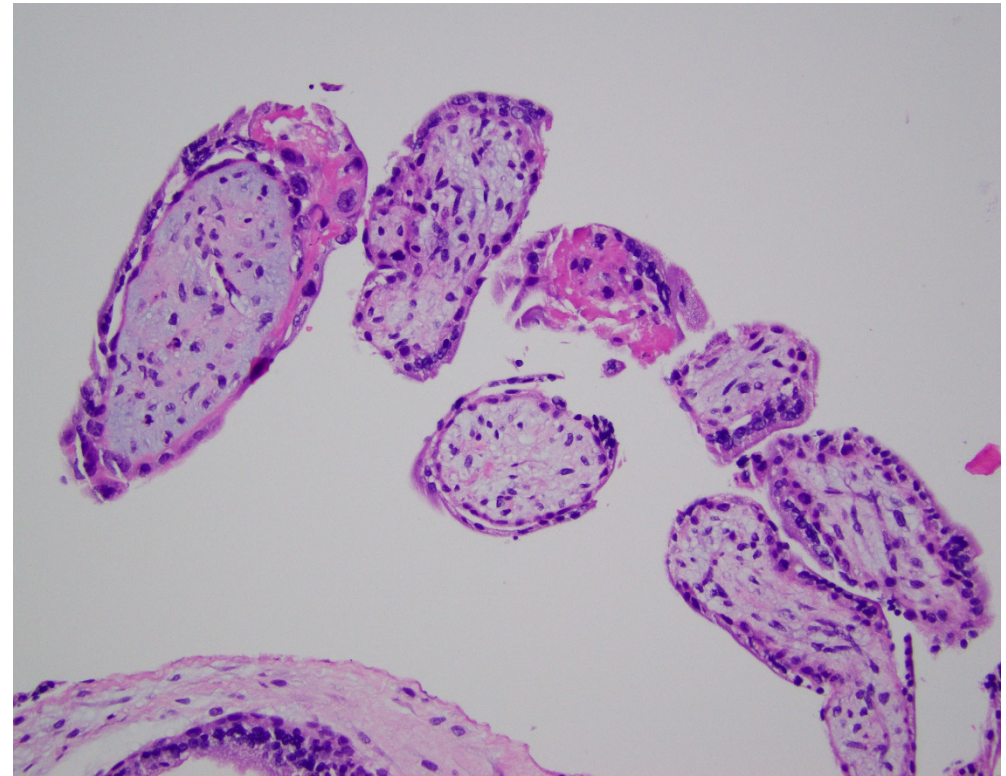


# Non-molar Triploidy (digynic - 2M1P)

**Not dimorphic**



**Villous sclerosis, without cavitation**



# Determination of Ploidy for PHM – The Solution, A Helping Hand or Unnecessary?

## Methods

- Karyotype
- Flow cytometry – DNA analysis
- In situ hybridization
- Microsatellite “fingerprinting”
- Microarray
- Whole genome/exome sequencing

## Issues

- Requires living fetal cells
- Needs whole nuclei
- Inconclusive with 1 chromosome
- Complicated without parental DNA
- Bioinformatics
- Bioinformatics
  
- All:
  - Instrumentation, Cost, & Time
  - Parent Of Origin
  - Data you didn't seek or on your radar

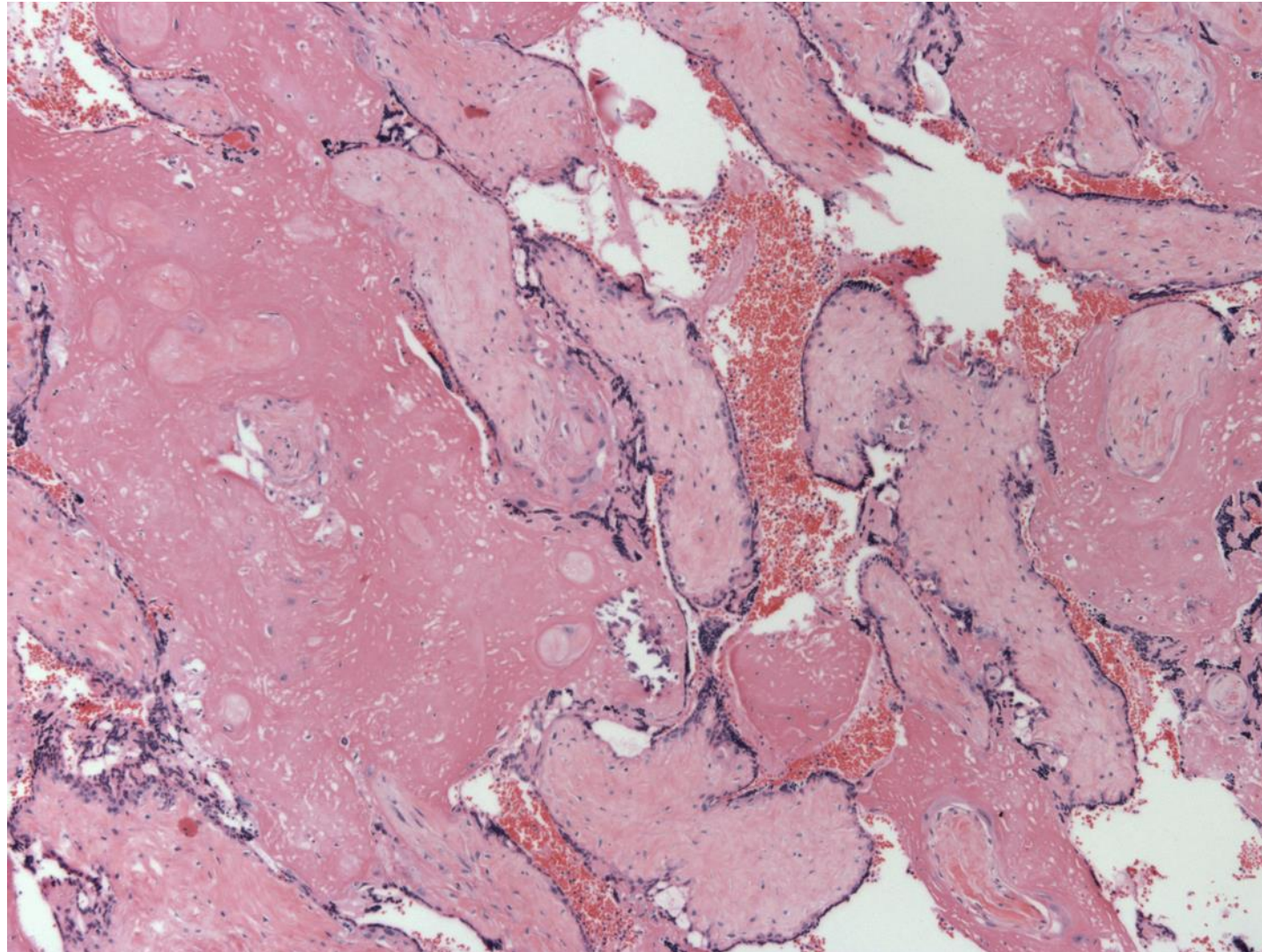


# Do you really need copy number analysis?

- **RESULTS:**

- 260 cases of PHMs
- Blinded pathologic review showed sensitivity of 88% and specificity of 95% in diagnosing PHMs with histology alone vs. histology with ancillary testing.
- 100% of PHMs (n=56) expressed p57 staining and 100% of CHMs (n=57) had absent staining
- 145 subjects had clinical follow-up data
  - 16 diagnosed with GTD (by serum hCG).
  - 13 cases of cases of clinical GTN histologically resembling ***“retained placenta”***
  - Only 2 had adverse outcomes (ETT and ChorioCA)

“Persistent” PHM looks like “funny” retained POC





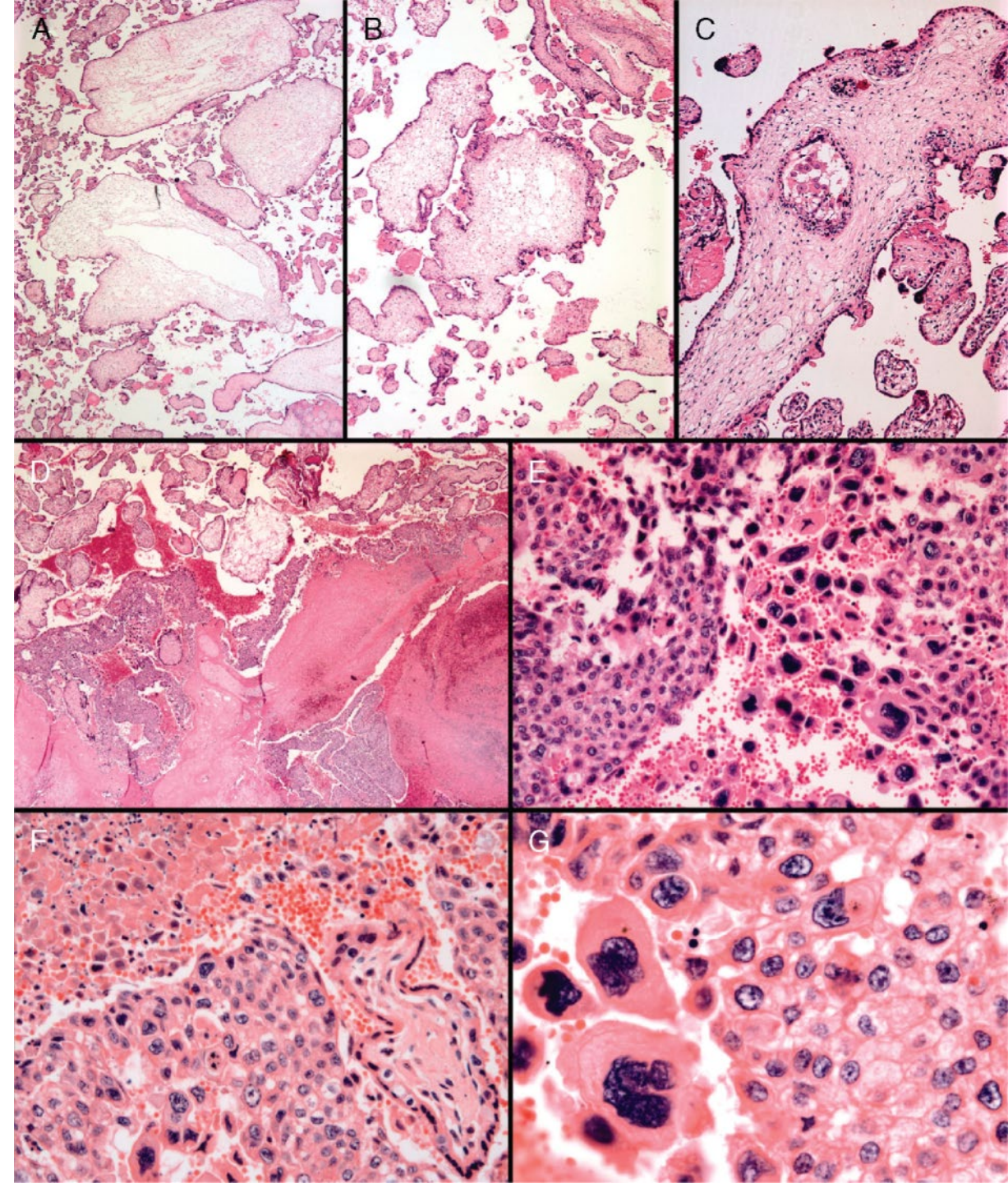
# The exception to the rule

*International Journal of Gynecological Pathology*  
27:247–251, Lippincott Williams & Wilkins, Baltimore  
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## Case Report

### Intraplacental Choriocarcinoma Arising in a Second Trimester Placenta With Partial Hydatidiform Mole

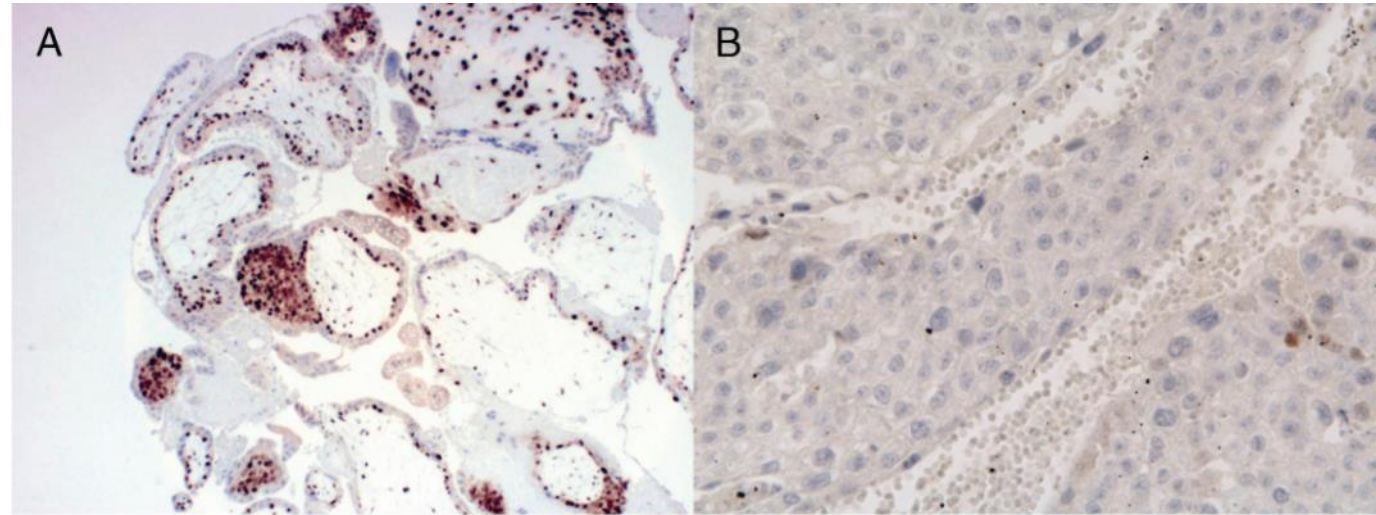
Fabiola Medeiros, M.D., Michael J. Callahan, M.D., Julia A. Elvin, M.D., Ph.D.,  
David M. Dorfman, M.D., Ph.D., Ross S. Berkowitz, M.D., and Bradley J. Quade, M.D., Ph.D.



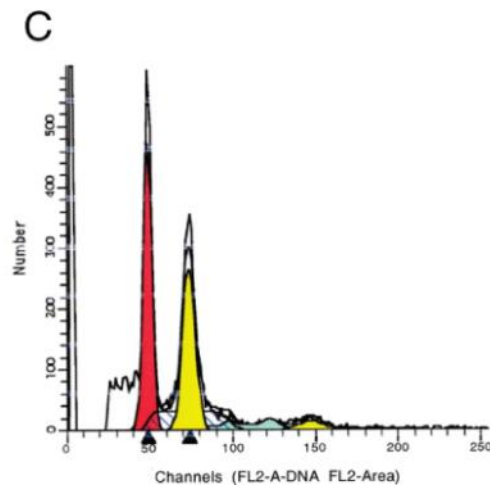


# Partial Hydatidiform Mole

P57 expression retained in PHM (A) and lost in ChorioCA (B)

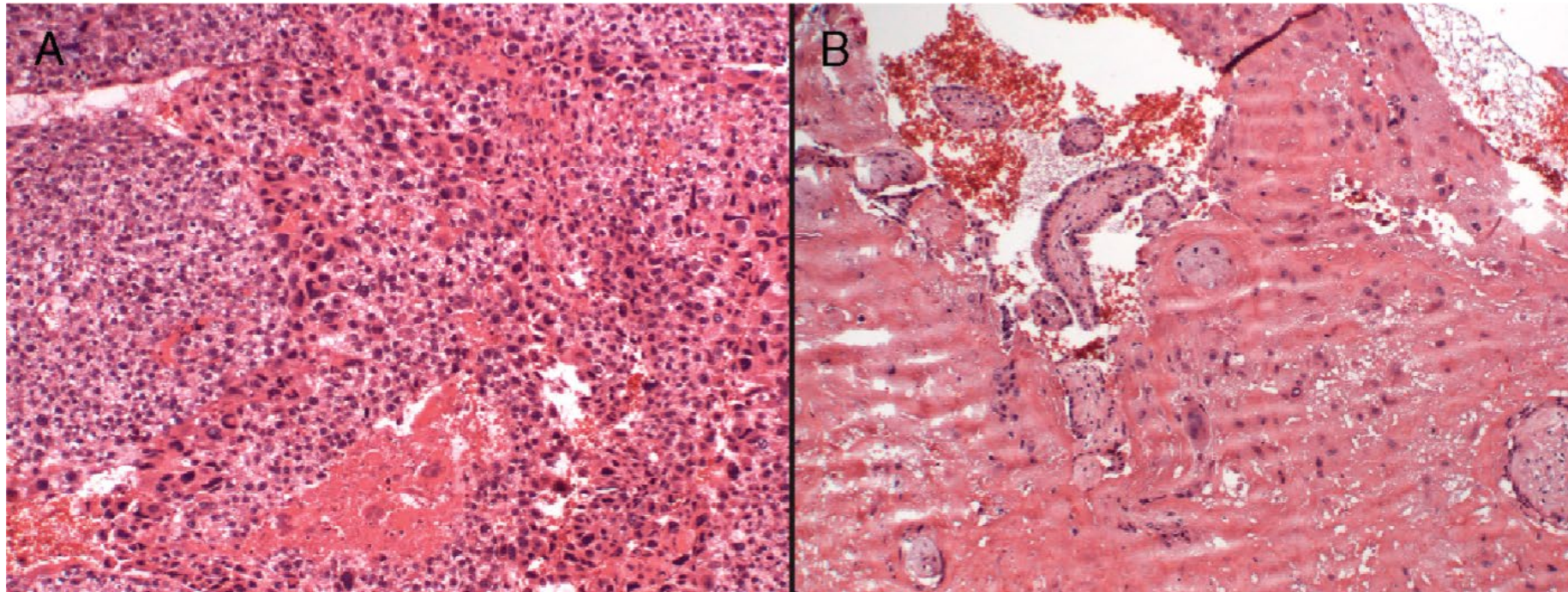


Triploid gestation (yellow peak) in placental





# Metastasis From Intraplacental Choriocarcinoma Arising In Partial Mole



*Diagnosing  
Possible Molar  
Gestations in  
General  
Surgical  
Pathology  
Practice*



The Scream by Edvard Munch - NPR



# A Practical Approach to Molar Pregnancies

- **Absence of villi does not exclude molar gestation... but it moves it down the list**
- **Risk of Persistent GTD (including recurrence or metastasis)**
  - CHD >> PHM (~20% v. ~2%, but metastatic risk closer to normal gestations for PHM)
- **Strategy should focus on finding every CHM**
  - Use p57 liberally...it works well, is cheap, and adds little delay
  - Ask for help if the p57 doesn't make sense (you may have found a chimeric or mosaic gestation with the potential to behave like a CHD biologically)
  - Use genetic information when you have it, after excluding a CHM
    - Some aneusomies (e.g., +13, +11, +21) are excellent mimics of PHM
    - Triploidy  $\neq$  PHM... but 3n plus dysmorphic villi (suggestive histology) is sufficient, in my opinion
    - I don't wait for karyotype or microarray in most cases (volume too high), but I add my name to cytogenetic report distribution and check it later
    - Non-canonical p57 might be twin or chimera/mosaic



**Timely Topic –**

**Maternal &  
Placental SARS-  
CoV-2**



# Fast Breaking Story

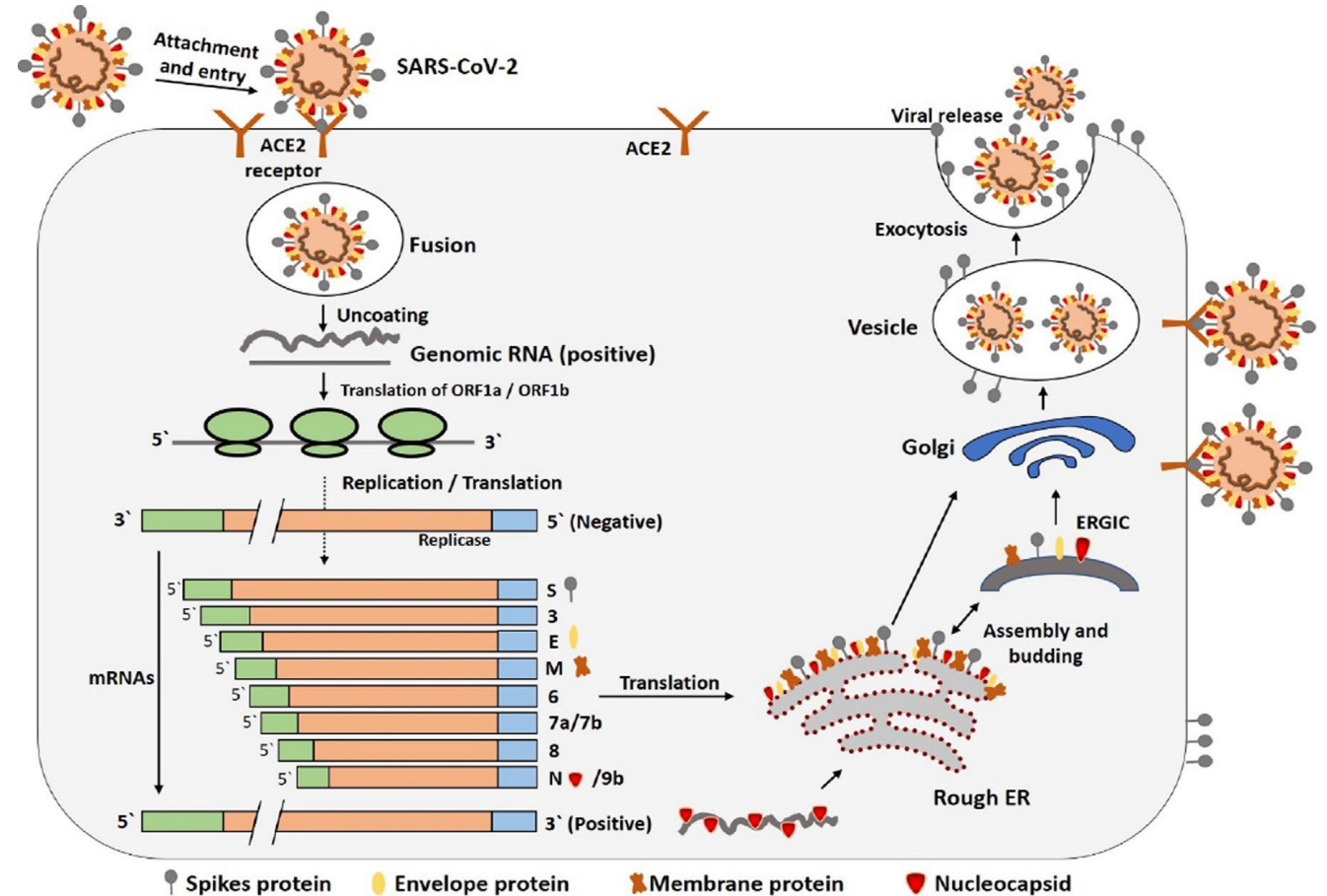
- From non-traditional outlets
  - Public (news) media
  - Social media
  - Professional list serves/bulletin boards, word-of-mouth
- Pre-publication release (not peer reviewed)...
  - the early standard practice for pandemic



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

**Fig. 3. The life cycle of SARS-CoV-2 in host cells;** begins its life cycle when S protein binds to the cellular receptor ACE2. After receptor binding, the conformation change in the S protein facilitates viral envelope fusion with the cell membrane through the endosomal pathway. Then SARS-CoV-2 releases RNA into the host cell. Genome RNA is translated into viral replicase polyproteins pp1a and 1ab, which are then cleaved into small products by viral proteinases. The polymerase produces a series of subgenomic mRNAs by discontinuous transcription and finally translated into relevant viral proteins. Viral proteins and genome RNA are subsequently assembled into virions in the ER and Golgi and then transported via vesicles and released out of the cell. ACE2, angiotensin-converting enzyme 2; ER, endoplasmic reticulum; ERGIC, ER–Golgi intermediate compartment.



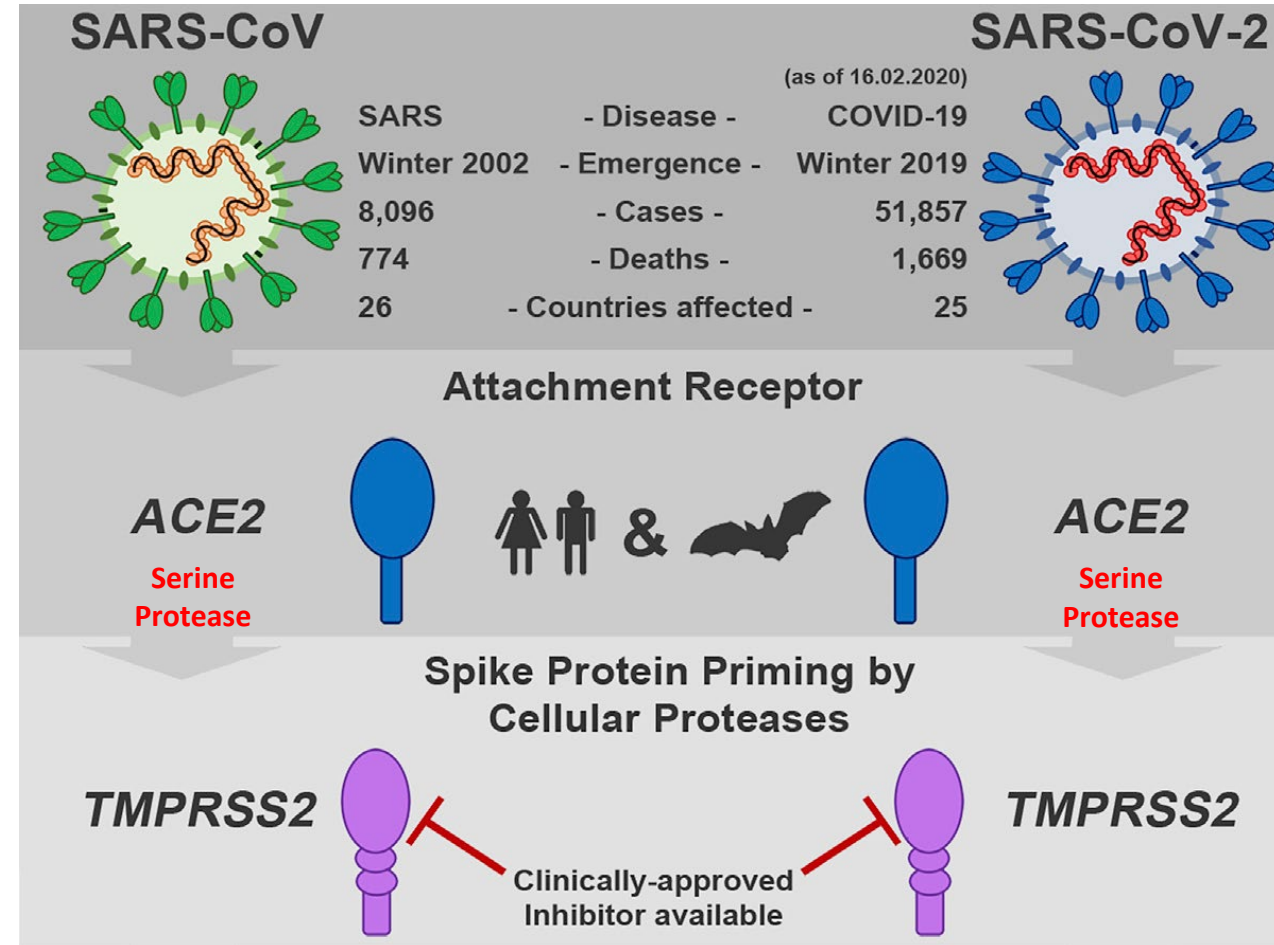


# SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor

Hoffmann et al., 2020, Cell 181, 271–280

April 16, 2020 © 2020 Elsevier Inc.

<https://doi.org/10.1016/j.cell.2020.02.052>



Transmission to  
the fetus and  
neonate:

How often and  
by what  
mechanism(s)?

- In utero – transplacental
- Intra-partem – vaginal contact
- Post-partem – breast milk



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<https://doi.org/10.5858/arpa.2020-0901-SA>

**Early Online Release**

**An Analysis of 38 Pregnant Women with COVID-19, Their Newborn Infants, and Maternal-Fetal Transmission of SARS-CoV-2: Maternal Coronavirus Infections and Pregnancy Outcomes**

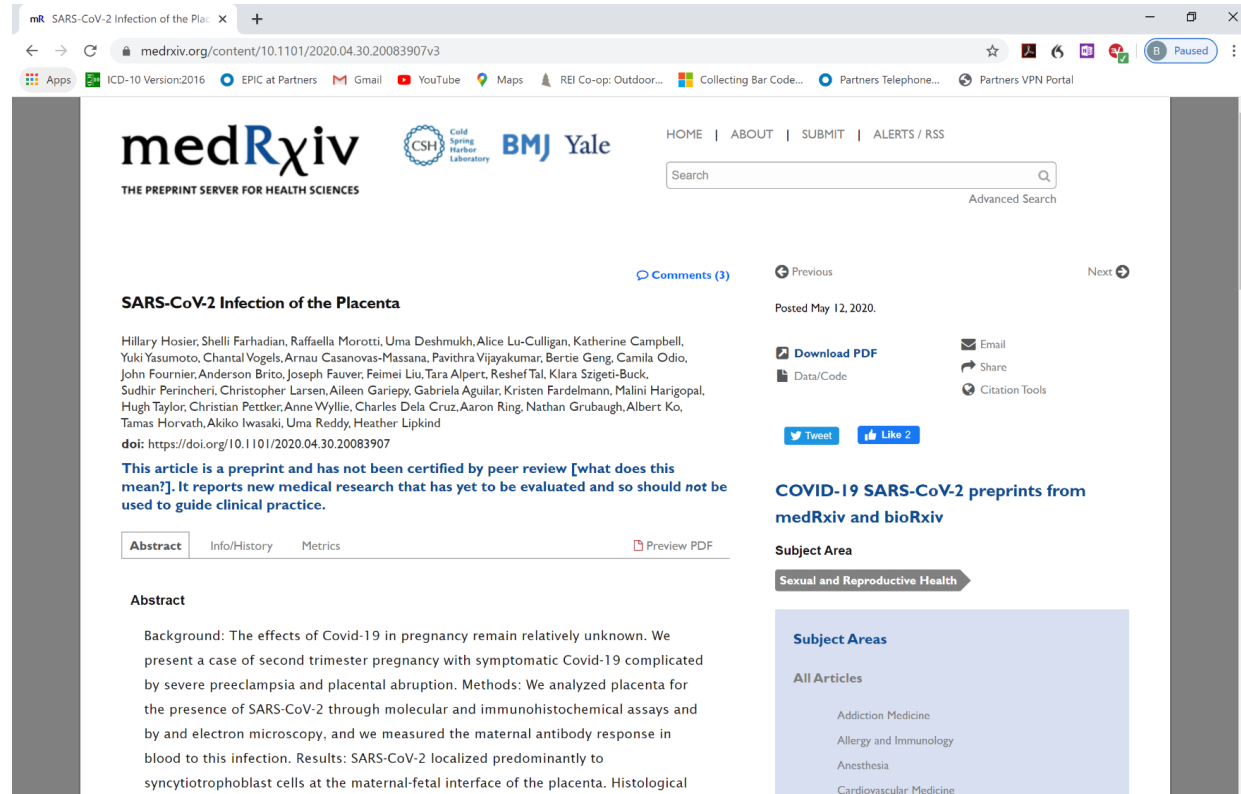
David A. Schwartz, MD, MS Hyg

The author has no relevant financial interest in the products or companies described in this article.

Corresponding author: David A. Schwartz, MD, MS Hyg, Department of Pathology, Medical College of Georgia, Augusta University, 1950 Grace Arbor Court, Atlanta, GA 30329 (email: [davidalanschwartz@gmail.com](mailto:davidalanschwartz@gmail.com)).

The emergence of a novel coronavirus, termed SARS-CoV-2, and the potentially life-threatening respiratory disease that it can produce, COVID-19, has rapidly spread across the globe creating a massive public health problem. Previous epidemics of many emerging viral infections have typically resulted in poor obstetrical outcomes including maternal morbidity and mortality, maternal-fetal transmission of the virus, and perinatal infections and death. This communication reviews the effects of two previous coronavirus infections - severe acute respiratory syndrome (SARS) caused by SARS-CoV and Middle East respiratory syndrome (MERS) caused by MERS-CoV - on pregnancy outcomes. In addition, it analyzes literature describing 38 pregnant women with COVID-19 and their newborns in China to assess the effects of SARS-CoV-2 on the mothers and infants including clinical, laboratory and virologic data, and the transmissibility of the virus from mother to fetus. This analysis reveals that unlike coronavirus infections of pregnant women caused by SARS and MERS, in these 38 pregnant women COVID-19 did not lead to maternal deaths. Importantly, and similar to pregnancies with SARS and MERS, there were no confirmed cases of intrauterine transmission of SARS-CoV-2 from mothers with COVID-19 to their fetuses. All neonatal specimens tested, including in some cases placentas, were negative by rt-PCR for SARS-CoV-2. At this point in the global pandemic of COVID-19 infection there is no evidence that SARS-CoV-2 undergoes intrauterine or transplacental transmission from infected pregnant women to their fetuses. Analysis of additional cases is necessary to determine if this remains true.

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medRxiv SARS-CoV-2 Infection of the Placenta

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CSH Cold Spring Harbor Laboratory BMJ Yale

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**Subject Areas**

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- Allergy and Immunology
- Anesthesia
- Cardiovascular Medicine

**SARS-CoV-2 Infection of the Placenta**

Hillary Hosier, Shelli Farhadian, Raffaella Morotti, Uma Deshmukh, Alice Lu-Culligan, Katherine Campbell, Yuki Yasumoto, Chantal Vogels, Arnau Casanovas-Massana, Pavithra Vijayakumar, Bertie Geng, Camila Odio, John Fournier, Anderson Brito, Joseph Fauver, Feimei Liu, Tara Alpert, Reshef Tal, Klara Szigeti-Buck, Sudhir Perincheri, Christopher Larsen, Aileen Gariepy, Gabriela Aguilar, Kristen Fardelmann, Malini Harigopal, Hugh Taylor, Christian Pettker, Anne Wyllie, Charles Dela Cruz, Aaron Ring, Nathan Grubaugh, Albert Ko, Tamas Horvath, Akiko Iwasaki, Uma Reddy, Heather Lipkind

doi: <https://doi.org/10.1101/2020.04.30.20083907>

**This article is a preprint and has not been certified by peer review [what does this mean?]. It reports new medical research that has yet to be evaluated and so should not be used to guide clinical practice.**

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**Abstract**

Background: The effects of Covid-19 in pregnancy remain relatively unknown. We present a case of second trimester pregnancy with symptomatic Covid-19 complicated by severe preeclampsia and placental abruption. Methods: We analyzed placenta for the presence of SARS-CoV-2 through molecular and immunohistochemical assays and by electron microscopy, and we measured the maternal antibody response in blood to this infection. Results: SARS-CoV-2 localized predominantly to syncytiotrophoblast cells at the maternal-fetal interface of the placenta. Histological

# New England Study

Modern Pathology (2020) 33:2092–2103  
<https://doi.org/10.1038/s41379-020-0639-4>



ARTICLE



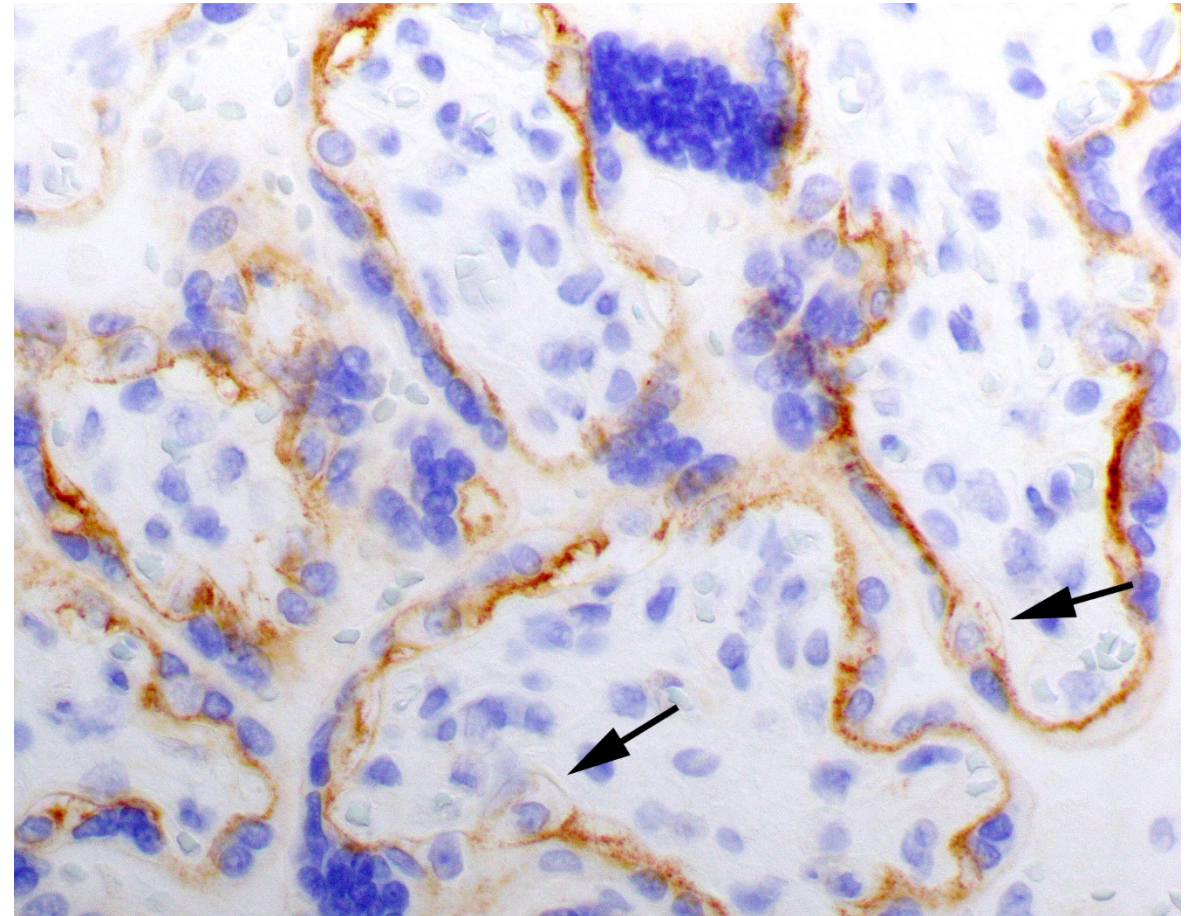
## SARS-CoV-2 can infect the placenta and is not associated with specific placental histopathology: a series of 19 placentas from COVID-19-positive mothers

Jonathon L. Hecht<sup>1</sup> · Bradley Quade<sup>2</sup> · Vikram Deshpande<sup>3</sup> · Mari Mino-Kenudson<sup>3</sup> · David T. Ting<sup>4,5</sup> · Niyati Desai<sup>4</sup> · Beata Dygulska<sup>6</sup> · Taryn Heyman<sup>7</sup> · Carolyn Salafia<sup>8</sup> · Dejun Shen<sup>9</sup> · Sara V. Bates<sup>10</sup> · Drucilla J. Roberts<sup>3</sup>

Received: 19 June 2020 / Revised: 24 July 2020 / Accepted: 24 July 2020 / Published online: 2 August 2020  
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- 19 COVID-19 exposed placentas
- ACE2 in syncytiotrophoblasts (ST) with polarized distribution skewed to **basal** side
- TMPRSS2 only present weakly in the villous endothelium and rarely in the ST
- Combined expression pattern may limit infection
- 2/19 infected placenta (“cherry picked”), no specific pathology

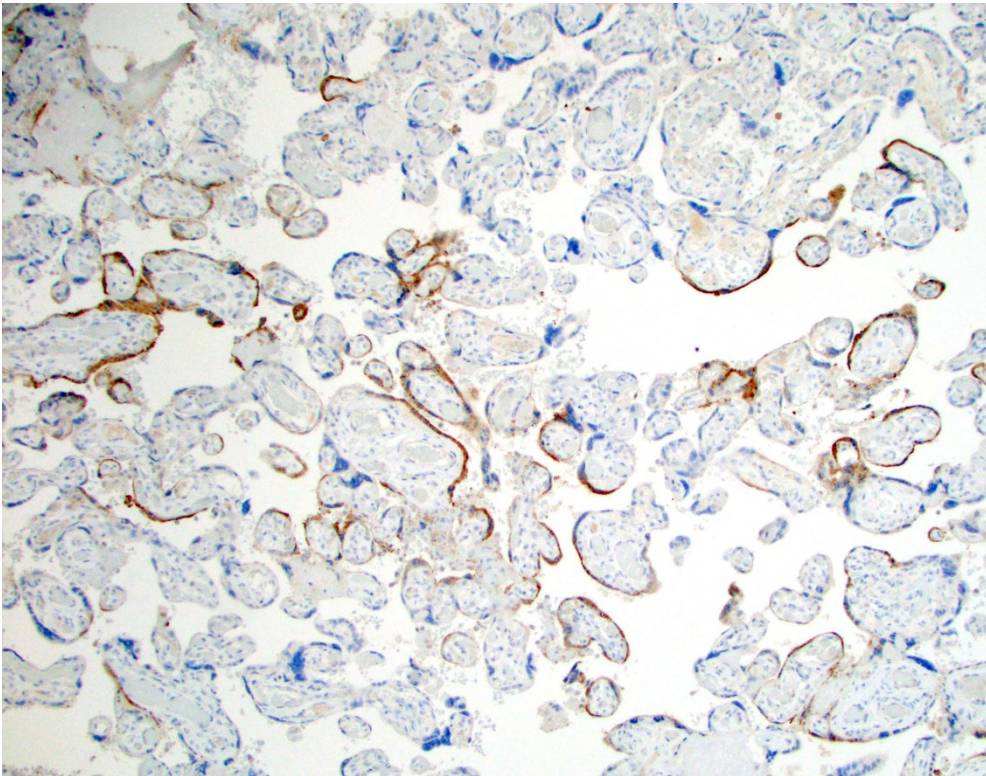
## ACE2



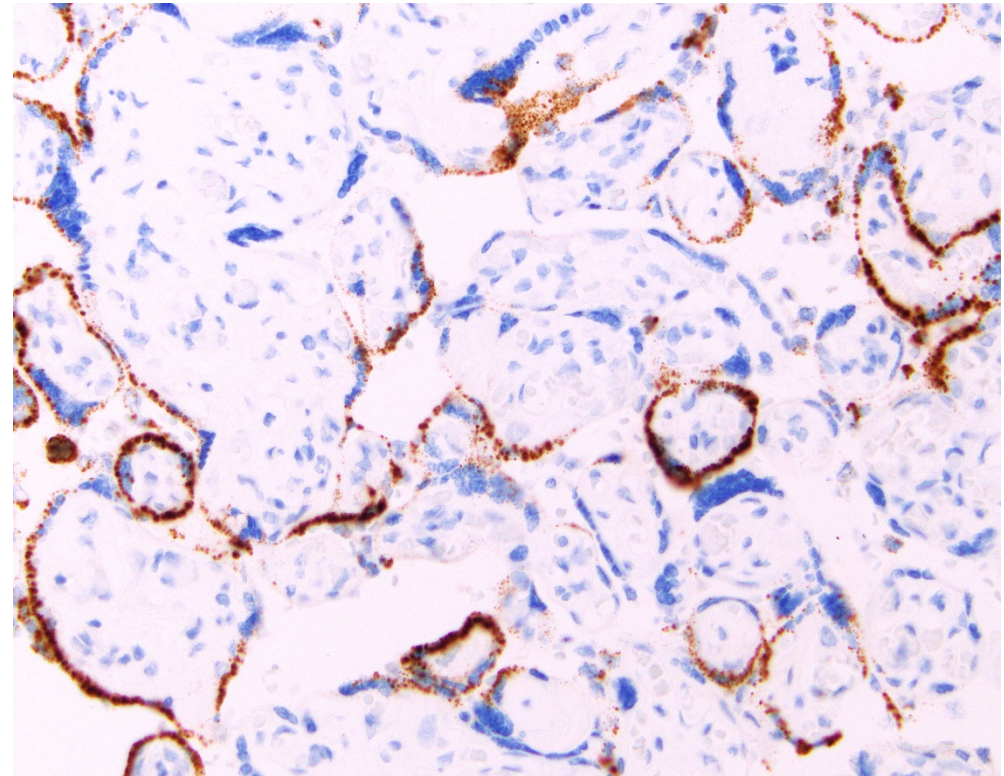


# Accumulation of More Case: Mother and Baby both COVID-19+

**Spike protein IHC**



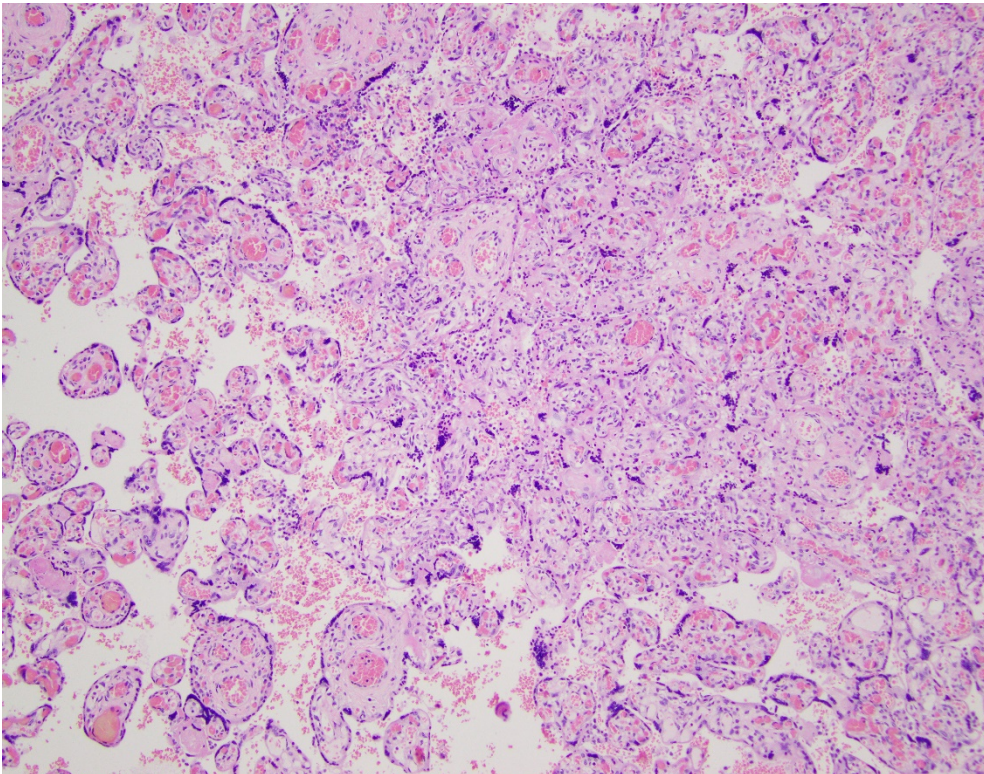
**Spike protein mRNA ISH**



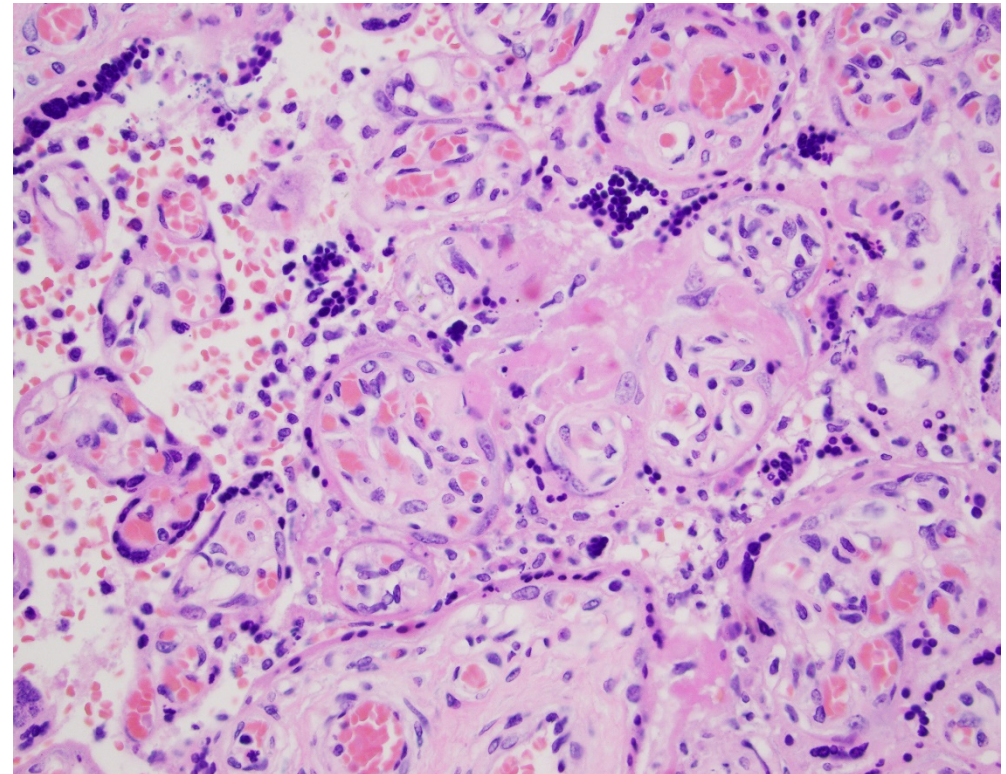


# COVID-19 Placentitis

**Various Sized Clusters Of Aggregated Villi**



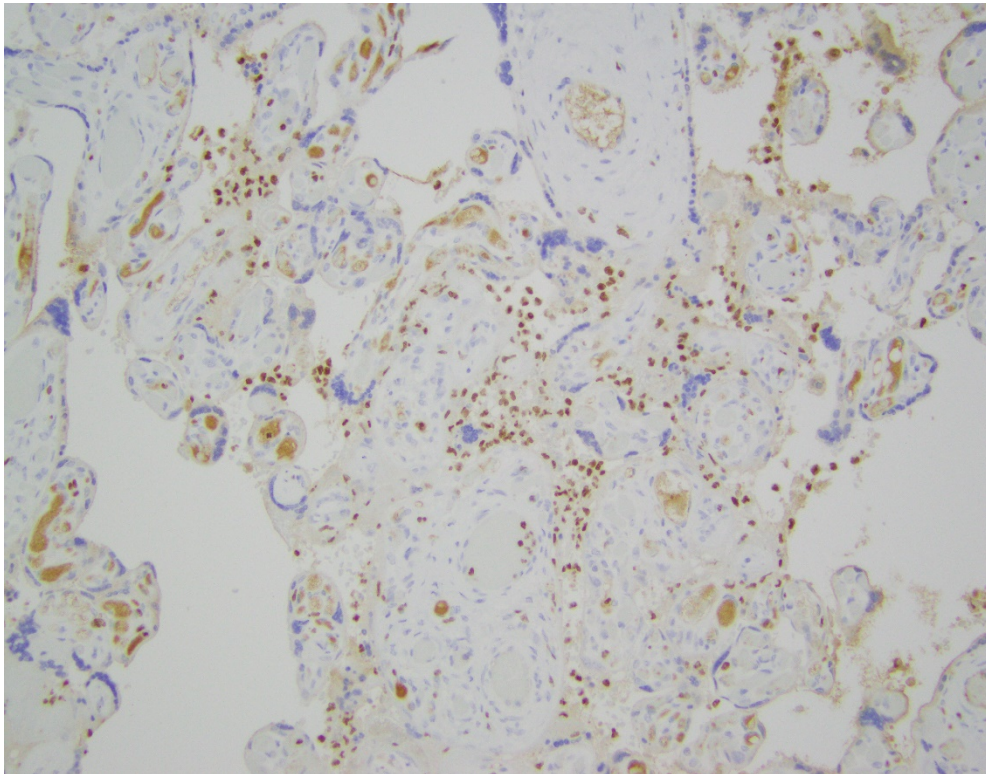
**Chronic Histiocytic Intervillositis & Early Placental Infarction/Intervillous Fibrin**



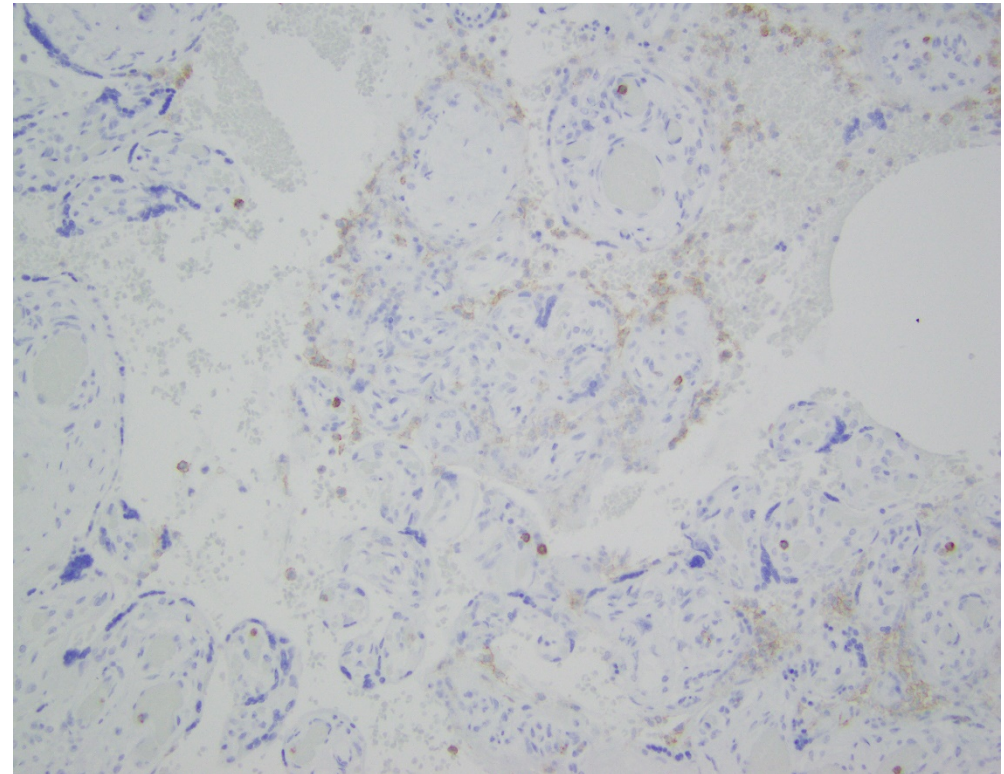


# COVID-19 Placentitis As Variant of Chronic Histiocytic Intervillositis

**Macrophages (PU.1)**



**Lymphocytes (LCA/CD45)**



# Take Away Messages

- **Placental infection uncommon** in most CoVID-19(+) mothers
- Mechanism (and clinical implications) of placental infection partially understood
  - **Maternal viremia low**
  - **ACE2** present in syncytiotrophoblasts...mostly on stromal side...and very low **TMPRSS2**, potentially limiting placental infection
- Role of ancillary testing for placenta not established...but **Anti-Spike IHC** likely to be helpful
- BOLO for **emerging pattern of histopathology** when SARS-CoV-2 infection present
  - **Atypical chronic histiocytic intervillitis** with
    - Mixed inflammatory infiltrate with PMN and macrophages
    - Necrosis / infarction
    - Perivillous fibrin deposition





KEEP  
CALM  
AND  
WASH  
YOUR  
HANDS





# Acknowledgements

- BWH / New England Trophoblast Disease Center
  - Ross Berkowitz
  - Neil Horowitz
  - Kevin Elias
  - Fabiola Medeiros
  - Rinda Soong
- Harvard
  - Dru Roberts
  - Johnathan Hecht





Thank You and Best Wishes for the New Year